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(54) Title: MYCOBACTERIUM TUBERCULOSIS GENES ENCODING PROTEIN ANTIGENS

(57) Abstract

Mycobacterium tuberculosis genes encoding five immunologically relevant proteins have been isolated by systematically screening a lambda gt11 recombinant DNA expression library with a collection of murine monoclonal antibodies dited against protein antigens of this pathogen. One of the M. tuberculosis antigens, a 65kD protein, has been shown to have determinants common to M. tuberculosis and M. leprae. In addition, genes encoding proteins of other mycobacteria (M. africanum, M. smegmatis, M. bovis BCG and M. avium) have been isolated. Isolation and characterization of genes encoding major protein antigens of M. tuberculosis make it possible to develop reagents useful in the diagnosis, prevention and treatment of tuberculosis. They can be used, for example, in the development of skin tests, serodiagnostic tests and reines specific for tuberculosis.

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MYCOBACTERIUM TUBERCULOSIS GENES AND ENCODING PROTEIN ANTIGENS

Description

Background

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Tuberculosis was the major cause of infectious mortality in Europe and the United States in the 19th and early 20th centuries. Dubos, R. and J. Dubos, The White Plague: Tuberculosis, Man and Society, Little Brown & Co., Boston, MA, (1952).

Today, it remains a significant global health

Today, it remains a significant global health problem.

For example, in the United States there are over 20,000 new cases of tuberculosis diagnosed annually. In addition, the steadily declining incidence of tuberculosis evident in preceding years appears to have changed course, reaching a plateau in 1985 and showing an increase in the first half of 1986. Centers for Disease Control, Morbidity/Mortality, Weekly Report, 34:774 (1986); and Centers for Disease Control, Morbidity/Mortality, Weekly Report, 35:774 (1986).

Worldwide, tuberculosis remains widespread and constitutes a health problem of major proportions, particularly in developing countries. The World Health Organization estimates that there are ten million new cases of active tuberculosis per year and an annual mortality of approximately three

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million. Joint International Union Against Tuberculosis and World Health Organization Study Group, <u>Tubercle</u>, 63:157-169 (1982).

Tuberculosis is caused by Mycobacterium (M.) 05 tuberculosis or Mycobacterium (M.) bovis, which are the 'tubercle bacilli' of the family Mycobacteriaceae. M. bovis is a species which causes tuberculosis in cattle and is transmissible to humans and other animals, in whom it causes tuberculosis. present, nearly all tuberculosis is caused by 1 û respiratory infection with M. tuberculosis. Infection may be asymptomatic in some, but in other individuals, it produces pulmonary lesions which lead to severe debilitation or death. Resistance to 15 tuberculosis is provided by cell-mediated immune mechanisms.

Mycobacteria are aerobic, acid-fast, non-sporeforming, non-motile bacili with high lipid contents
and slow generation times. M. leprae is the etiologic agent of leprosy and, among the other mycobacteria, the only major pathogen. Bloom, B.R. and
T. Godal, Review of Infectious Diseases, 5:765-780
(1983). However, other mycobacterial species are
capable of causing disease. Wallace, R.J. et.al.,
Review of Infectious Diseases, 5:657-679 (1984).
M.avium, for example, causes tuberculosis in fowl
and in other birds. Members of the M.
Avium-intracellularae complex have become important
pathogens among individuals with acquired immunodeficiency syndrome (AIDS). Certain groups of

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individuals with AIDS have a markedly increased incidence of tuberculosis-as well. Pitchenik, A.E. et. al., Annals of Internal Medicine, 101:641-645 (1984).

Diagnostic and immunoprophylatic measures for mycobacterial diseases have changed little in the past half century. Tuberculin, developed by Koch as a cure for tuberculosis in the late 1800s, is an M. tuberculosis filtrate of complex and poorly-defined composition. It is used as a skin test antigen to detect prior exposure to the bacillus. Enrichment of the protein fraction of this material in the 1930's produced the purified protein derivative (PPD) which is still used to diagnose exposure to tuberculosis. Seibert, F.M. et.al., American Review of Tuberculosis, 30(Suppl.):705-778 (1934). Its usefulness is limited, however, by its lack of specificity and its inability to distinguish active disease from prior sensitization by contact with M. tuberculosis or cross-sensitization to other mycobacteria. Young, R.A. and R.W. Davis, Proceedings of the National Academy of Sciences, USA, 80:194-1198 (1983).

Bacille Calmette Guerin (BCG), an avirulent

strain of M. bovis, has been used widely as a live
vaccine against tuberculosis for over 50 years.

Calmette, A., C. et.al., Bulletin of the Academy of

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Medicine Paris, 91:787-796 (1924). During that time, numerous studies have shown that BCG has protective efficacy against tuberculosis. studies are reviewed by F. Luelmo in American Review of Respiratory Diseases, 125(pt. 2):70-72 (1982). However, more recently, a major trial of BCG in India indicated that such a vaccine was not protective against tuberculosis in this setting. World Health Organization WHO Technical Report Series, 651 (1980). Presently available approaches to diagnosing, preventing and treating tuberculosis are limited in their effectiveness and must be improved if a solution is to be found for the important public health problem tuberculosis represents worldwide.

Summary of the Invention

The present invention is based on the isolation of genes encoding immunogenic protein antigens of the tubercle bacillus Mycobacterium tuberculosis (M. tuberculosis). Genes encoding such protein antigens have been isolated from a recombinant DNA expression library of M. tuberculosis DNA. Genes encoding proteins of four additional mycobacteria have also been isolated and restriction maps produced.

In particular, genes encoding five immunodominant protein antigens of the tuberculosis bacillus (i.e., those M. tuberculosis proteins of molecular weight 12,000 daltons (12kD), 14kD, 19kD, 65kD and 71kD have been isolated by probing a lambda gt11 expression library of M. tuberculosis DNA with

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monoclonal antibodies directed against \underline{M} . tuberculosis-specific antigens.

Recombinant DNA clones producing the specific antigenic determinants recognized by the monoclonal antigens were also isolated in this manner. DNA from such recombinant lambda gtl1 clones was mapped with restriction endonucleases; the restriction maps for genes encoding the five immunodominant protein antigens (i.e., genes encoding the 12kD, 14kD, 19kD, 65kD and 71kD proteins) were deduced. The nucleotide sequence of three of the genes have been determined and, in each case, the amino acid sequence of the encoded protein has been deduced.

Brief Description of the Drawings

Figure 1 shows restriction maps of M. tuberculosis DNA. Recombinant DNA clones isolated with
monoclonal antibodies directed against the 12kD,
14kD, 19kD, 65kD and 71kD protein antigens were
mapped with restriction endonucleases. The insert
DNA endpoints are designated left (L) or right (R)
in relation to lac Z transcripts which traverse the
insert from right to left. Restriction sites are
represented as follows: A, Sal I; B, BamHI; E,
EcoRI; G, BglII; K, KpnI; P, PvuI; S, SacI; X, XhoI.

Figure 2 shows arrays of antigens from M.

tuberculosis recombinant DNA clones probed with
rabbit hyperimmune serum. The code of the recombinant DNA clones shown on the numbers filter is: 1,
Y3275; 2, Y3274; 3, Y3279; 4, Y3277; 5, Y3247; 6,
Y3272; 7, Y3150; 8, Y3254; 9, Y3147; 10, Y3163; 11,

Y3179; 12, Y3191; 13, Y3252; 14, Y3178; 15, Y3180; 16, Y3143; 17, lambda gtll. Clones 1, 5, 6, 7, 9 and 16 are M. tuberculosis recombinants described in the following section. Clones 10, 11, 14 and 15 are M. leprae recombinants expressing epitopes of the 18kD, 28kD, 36kD and 65kD antigens, respectively. Clones 2, 3, 4, 8, 12, 13 are uncharacterized recombinants from the lambda gtll M. tuberculosis and M. leprae libraries. Clone 17 is a non-recombinant lambda gtll control.

Figure 3 shows arrays of recombinant mycobacterial antigens probed with monoclonal antibodies to assess the extent of cross-reactivity between recombinant protein antigen of M. tuberculosis and of M. leprae. The array of clones is identical to that shown in Figure 2. Antibody probes and the antigen sizes recognized are: 1, IT-11 (71kD); 2, IT-31 (65kD); 3, IT-16 (19kD); 4, IT-1 (14kD); 5, IT-3 (12kD).

Figure 4 shows restriction maps of DNA encoding four proteins (71kD, 65kD, 19kD and 14kD) of M.

tuberculosis and four proteins (71kD, 65kD, 19kD and 14kD) of M. bovis BCG. Restriction sites are represented as follows: A, AatII; B, BamHl; C,

BclI; D, DraIII; E=EcoRI; G, BglII; H, HinfI; K, KpnI; P, PstI; S, SalI; V, PvuI and X, XhoI.

Figure 5 is a comparison of restriction maps of the gene encoding the 65kD protein of 6 mycobacteria (M. leprae, M. tuberculosis, M. africanum, M. bovis

30 BCG, M. smegmatis, M. avium). Restriction sites are

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as follows: B, BamHl; K, KpnI; N, SacI; P, PvuI; S, SalI; X, XhoI.

Figure 6 is the nucleotide sequence of the region containing the <u>M. tuberculosis</u> 19kD gene. The deduced amino acid sequence of the encoded protein is also represented (protein start position, nucleotide 1110; protein stop position, nucleotide 1586).

Figure 7 is the nucleotide sequence of the region containing the M. tuberculosis 71kD gene and the deduced amino acid sequence of the encoded protein.

Figure 8 is the nucleotide sequence of the region containing the <u>M. tuberculosis</u> 65kD gene.

The deduced amino acid sequences of the two long open reading frames are presented in one letter code over (540) or under (517) the appropriate triplets.

Detailed Description of the Invention

The invention described herein is based on the isolation of genes encoding immunogenic protein antigens of the bacillus M. tuberculosis, which is the major etiologic agent of tuberculosis. In particular, it is based on the isolation, using monoclonal antibodies directed against M.

- tuberculosis-specific antigens, of genes encoding five immunogenic protein antigens of the tuberculosis bacillus; these five antigens are immunodominant. Immunogenic antigens are those which elicit a response from the immune system.
- 30 Immunodominant protein antigens are immunogenic

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antigens against which the immune system directs a significant portion of its response. Genes encoding M. tuberculosis antigens of molecular weight 12,000 daltons (12kD), 14kD, 19kD, 65kD and 71kD were isolated in this manner.

Isolation and characterization of major protein antigens of <u>M. tuberculosis</u>, as described herein, make it possible to develop more effective tools for the prevention, diagnosis, and treatment of tuberculosis. Identification and isolation of genes encoding five immuncdominant <u>M. tuberculosis</u> protein antigens, as well as of the five protein antigens, are described below; uses of the genes and encoded products are also described.

M. bovis BCG DNA clones were also isolated for the genes encoding the 71kD, 65kD, 19kD and 14kD proteins. In order to compare M. bovis BCG and M. tuberculosis genes encoding proteins of similar molecular weight, restriction endonuclease maps were determined for DNA segments containing each of the genes. Restriction maps for each of these genes is represented in Figure 4.

In addition, DNA clones were isolated for the genes encoding the 65kD protein from M. africanum,

M. smegmatis and M. avium. Restriction endonuclease maps were determined for DNA segments containing each of these genes. The restriction maps for these genes, as well as for the genes encoding the 65kD protein of M. tuberculosis, M. bovis BCG and M.

leprae, are represented in Figure 5.

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I. Construction of a recombinant expression library of M. tuberculosis DNA

A recombinant DNA expression library of M.

tuberculosis DNA was constructed using lambda gtll.

The library was constructed with M. tuberculosis

genomic DNA fragments in such a way that all

protein-coding sequences would be represented and

expressed. Young, R.A., B.R. Bloom, C.M.

Grosskinsky, J. Ivanyi, D. Thomas and R.W. Davis,

Proceedings of the National Academy of Sciences,

USA, 82:2583-2587 (1985).

Lambda gtll is a bacteriophage vector which is capable of driving the expression of foreign insert DNA with E. coli transcription and translation signals. Lambda gtll expresses the insert DNA as a fusion protein connected to the E. coli Betagalactosidase polypeptide. This approach ensures that the foreign DNA sequence will be efficiently transcribed and translated in E. coli. This approach is also useful in addressing the problem of the highly unstable nature of most foreign proteins; fusion proteins are often more resistant to proteolytic degradation than is the foreign polypeptide alone. Lambda gtll and the E. coli strain used (Y1090) have been described previously. Young, R.A. et al., Proceedings of the National Academy of Sciences, USA, 80:1194-1198 (1983); Young, R.A. and R.W. Davis, Science, 222:778-782 (1983). The teachings of these publications are incorporated herein by reference. The library constructed in this manner has a titer of 1x 10¹⁰ pfu/ml. and

contains approximately 40% recombinants with an average insert size of $4\overline{kB}$.

II. Screening of the lambda gtll M. tuberculosis library with antibody probes

Murine monoclonal antibodies to protein antigens of M. tuberculosis were used individually to
probe the M. tuberculosis recombinant DNA library.
This work is described below and with specific
reference to the 65kD antigen in the Exemplification. The antibodies used as probes and the sizes
of the antigens to which they bind are shown below.

		M. tuberculosis
	Antibody	<u>Antigen</u>
	IT-3	12kD
15	IT-20	14kD
	IT-19	19kD
	IT-27	19kD
	IT-17	23kD
	IT-29	23kD
20	: IT-15	. 38kD
	IT-21	38kD
	IT-23	38kD
	IT-13	65kD
	IT-31	65kD
25	IT-33	65kD
	IT-11	71kD

Engers, H.D. et al., <u>Infectious Immunology</u>, <u>51</u>:718-720 (1986).

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All monoclonal antibodies were used at approximately 1:200 to 1:300 dilution in 50mM Tris-HCl pH8/150 mM NaCl/.05% Tween 20.

Screening of the lambda gtll recombinant DNA library was performed as described by Young et al. in Proceedings of the National Academy of Sciences, USA, 82:2583-2587 (1985), the teachings of which are incorporated herein by reference. One modification was made in the method described by Young and co-workers: 1% bovine serum albumin was used in place of 20% fetal calf serum to decrease background.

Briefly, cloned lambda gtll recombinants were arrayed on lawns of E. coli Y1090. The phage were grown, antigen expression was induced and the antigens were blotted and probed with serum. Detection of signal-producing plaques was performed with a biotinylated secondary antibody system (Vectastain, Vector Laboratories, Burlingame, CA) or with an alkaline phosphatase conjugated secondary antibody system (Protoblot, Promega Biotec, Madison, WI), both used according to manufacturer's instructions. Signal-producing clones were isolated using antibodies directed against protein antigens of molecular weight 12kD, 14kD, 19kD and 65kD and 71kD. In each case, similar numbers of clones were isolated in screens of approximately 105 recombinant plagues. DNA clones encoding the 23kD and 38kD antigens could not be detected with these antibodies, possibly because the native epitope is modified or topographically complex, or because the

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antigen-antibody interaction is too weak to be recognized by current detection methods.

III. Probing of Arrays of lambda gtll DNA Clones with Antibody Probes

0.2 ml of a saturated culture of Y1090 was added to 2.5 ml of molten LB soft agar, poured onto 100 mm plates containing 1.5% LB agar and allowed to harden at room temperature for 10 min. phage plate stock containing approximately 1011 pfu/ml of the lambda gtll DNA clones of interest were placed into alternate wells of 96-well tissue culture plates. A multi-pronged transfer device was placed briefly in the wells containing phage and then touched lightly to the surface of the plate onto which the soft agar had been poured. plates were then incubated at 42°C for approximately 3 hours, at which point clear plaques approximately 5mm in diameter were visible. The plates were then overlayed with nitrocellulose filters saturated with 10mM isopropylthiogalactoside (IPTG) and incubated at 37°C for 3.5 hours. Subsequent processing of filters for detection of antigen was identical to the procedures described for screening of lambda

Immunoscreening of the lambda gtll library to isolate clones reactive with monoclonal antibodies specific for the 65kD antigen is described in the Exemplification.

gtll library with antibody probes.

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IV. Recombinant DNA Manipulation

DNA from recombinant lambda gtll clones was isolated and mapped with restriction endonucleases by standard techniques. Davis, R.W. et al., Advanced Bacterial Genetics: A Manual for Genetic

O5 Advanced Bacterial Genetics: A Manual for Genetic Engineering, Cold Spring Harbor (1980).

Figure 1 shows the genomic DNA restriction map deduced for each of the genes encoding the five M. tuberculosis antigens and illustrates how each of the cloned DNAs aligns with that map. All clones isolated with monoclonal antibodies directed against any single antigen align with a single genomic DNA segment. This indicates that all clones were isolated because they express the protein of interest rather than an unrelated polypeptide containing a similar or identical epitope. In addition, this result suggests that each antigen is the product of a single gene.

The orientation of each DNA insert in the

recombinant clones was determined by restriction
analysis. Only among the clones for the 65kD
antigen were the inserts found in both possible
orientations relative to the direction of lac 2
transcription in lambda gtll. This suggests that

this protein can be expressed in E. coli from
signals independent of those provided by lac 2.
Similar results have been obtained for recombinant
DNA clones encoding the 65kD antigens of M. bovis
and M. leprae. Thole, J.E.R. et al., Infectious
Immunology, 50:800-806 (1985); Young, R.A. et al.,

Nature, 316:450-452 (1985).

The nucleotide sequences of three regions of the <u>M. tuberculosis</u> DNA were determined: 1) the region containing the <u>M. tuberculosis</u> 19kD gene; 2) the region containing the <u>M. tuberculosis</u> 71kD gene; and 3) the region containing the 65kD gene. The three sequences are represented in Figures 6-8. Sequences were determined using standard techniques, which are described in the Exemplification.

V. Filter hybridization of Insert DNA

10 Arrays of lambda gtl1 clones were created as described above and incubated at 42° for 5 hours. The plates were then overlayed with nitrocellulose filters and placed at 4°C for 1 hour. Probe DNA was labelled with ³²P by nick translation. Filter hybridization was performed as described by Davis et 15 al. in Advanced Bacterial Genetics: A Manual for Genetic Engineering, Cold Spring Harbor (1980), the teachings of which are incorporated herein by reference. Hybridization conditions were as follows: 50% v/v formamide, 5x SSPE (1x SSPE is 0.18M NaCl, 10mM 20 Na, 5H, 5PO, 1mM Na, EDTA, pH 7.0), 1x Denhardt's solution (0.02% w/v Ficoll, 0.02% w/v polyvinylpyrrolidone, 0.02% w/v bovine serum albumin), 0.3% NaDodSO_A at 42°C for approximately 16 hours, followed by washing in 2x SSPE, 0.2% NaDodSO4 at 45°C. 25

VI. Recombinant Antigens Recognized by Rabbit Serum

The response of a second animal to an antigen preparation of \underline{M} . tuberculosis was assessed by

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examining the reactivity of rabbit anti-M. tuberculosis hyperimmune sera with recombinant antigens. Cloned lambda gtll recombinants were arrayed on lawns of E. coli and probed with the rabbit hyperimmune serum. Anti-M. tuberculosis hyperimmune serum, produced by repeated immunization of rabbits with M. tuberculosis H37Rv culture filtrate, was provided by J. Bennedsen (Statens Seruminstitut, Copenhagen, Denmark). These sera were used at 1:100 dilution.

These sera produced positive signals with lambda gtll clones encoding each of the five M. tuberculosis epitopes which had been isolated with murine monoclonal antibodies (Figure 2). Particularly strong signals were observed with the 65kD and 71kD antigens (Figure 2). These results demonstrate that mice and rabbits can mount an antibody response to the same protein antigens of M. tuberculosis.

Clones for the five M. tuberculosis antigens were detected at similar frequencies in the lambda gtll recombinant DNA library. Thus, the number and type of antigen-producing clones isolated with polyclonal serum antibodies should reflect the relative titer and deversity of the individual antibodies in this serum.

To determine whether any of the 5 M. tuberculosis antigens are relatively immunodominant in the rabbit humoral immune response to M. tuberculosis, the M. tuberculosis lambda gtll recombinant DNA library was screened with the rabbit serum. Forty signal-producing clones were isolated, arrayed on

lawns of E. coli Y1090 and probed with monoclonal antibodies directed against each of the 5 recombinant M. tuberculosis protein antigens. Remarkably, 17 of the 40 clones (43%) reacted strongly with at 05 least one of the four anti-65kD monoclonal antibodies tested. An additional six clones (15%) reacted strongly with the anti-17kD monoclonal antibody (IT-11). This indicates that a large proportion of the anti-M. tuberculosis antibody 10 present in the rabbit serum was directed against the 65kD antigen of M. tuberculosis and suggests that it is a dominant antigen for the rabbit humoral immune response. Seventeen of the clones did not react with any of the monoclonal antibodies tested, suggesting that the rabbit sera may identify M. 15 tuberculosis proteins not recognized by the murine antibodies.

VII. Antigenic Relatedness of M. tuberculosis and M. leprae Proteins

There is evidence that <u>M. tuberculosis</u> and <u>M. leprae</u> share immunologically important antigens. To assess this further, an investigation of the exact nature of the immunological relatedness among recombinant protein antigens of <u>M. tuberculosis</u> and <u>M. leprae</u> was conducted.

For each of five \underline{M} . <u>tuberculosis</u> and four \underline{M} . <u>leprae</u> protein antigens, a single recombinant DNA clone containing most or all of the gene of interest was used to express antigen in the following manner. The recombinant phage clones were arrayed on a lawn

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of <u>E</u>. <u>coli</u> Y1090, which was then grown and induced for antigen expression.

Antigen immobilized on nitrocellulose filters was then probed with 26 individual anti-M. tuberculosis and M. leprae monoclonal antibodies. Figure 3 shows the array of DNA clones used and the results obtained with the anti-M tuberculosis antibodies IT-1, IT-3, IT-11, IT-16, and IT-31, which recognized proteins of 14kD, 12kD, 71kD, 19kD and 65kD respectively. Table 1 details the full results of these cross-screening experiments, showing the reactivity of antigen expressed from individual recombinant DNA clones with each of the individual monoclonal antibodies. Clones were scored as positive only if the signal produced was clearly greater than the background signal produced by the non-recombinant lambda gtll clone included in each array.

TABLE 1

Reactivity of Monoclonal Antibodies with

Recombinant Protein Antigens

				-		ANG	CLONES			_	
•			R: 27	berculo	ate				H. le	DE3G	
ANTIBODIES .		12k0 Y3275	14KD Y3247	19kD Y3147	65kD Y3150	71XD Y3272	- 19t11	18kD Y3179	28kD Y3163	36XD Y3180	65kD Y3178
H. tube:	culosis										
12KD	IT-3	\odot	-	-	-	•	-	•	•	-	-
14kD	IT-1	-	①	-	-	-	,-	-	-	-	-
	IT-4	-	\odot	•	•	-	•	-	-	-	-
	IT-20	•	•	-	-	-	-	-	-	-	
19k0	IT-10	-	-	①	-	•	•	•	-	-	-
	IT-12	-	-	•	•	•	•	-	-	-	-
	IT-16	-	-	⊙	-	-	-	-	-	-	-
	IT-19	-	-	•	-	•	-	•	•	-	-
65kD	IT-13	-	-	-	⊙	•	•	-	-	-	-
	17-31	-	-	-	\odot	-	-	-	-	-	\odot
	17-33	-	-	-	\odot	-	•	-	•	-	\odot
71kD	IT-11	-	-	-	-	\odot	-	-,	-	-	•
M. lepra	<u>.e</u>										
18kD	L7-15	-	-	-	•	-	•	•	-	-	-
28kD	SA1.D2D	-	-	-	_	-	. •		O .		-
	SA1.BII	: -	. -	-	-	-	-	•	-	-	-
36kD	F47-9-1	-	-	-	-	-	-	•	-	\odot	-
	HLO4-A	-	-	-	-	* -	- ·		-	-	•
65kD	cI.I	-	-	-	\odot	-	-	-	-	-	\odot
	IIH9	-	•	-	\odot	-	-	-	-	-	\odot
	IIIE9	-	-	-	-	-	-	-	•	•	\odot
•	IICS	-	•	-	\odot	•	•	-	-	-	\odot
	IIIC8	-	•	-	-	-	•	-	-	•	\odot
	T2.3	•	-	-	\odot	-	-	-	•	•	\odot
	Y1-2	-	-	• .	\odot	-	•	-	•	-	0000000
	SA2.D7C	-	•	-	\odot	-	-		•	•	\odot
	HLJOA	\odot	\odot	\odot	Ō	⊙ ·	\odot	\odot	\odot	\odot	\odot

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Several conclusions can be drawn from the results shown in Table 1. Among the 11 monoclonal antibodies that recognize a 65kD antigen, 7 react with the 65kD protein from both mycobacteria (IT-31, C1.1, IIH9 (identical to IT-33), IIC8, T2.3, Y1-2, SA2.D7C), one antibody reacts only with the \underline{M} . tuberculosis 65kD protein (IT-13), and two antibodies react only with the \underline{M} . leprae 65kD protein (IIIE9 and IIIC8). One antibody, ML30A, cross-reacts with an antigen in \underline{E} . \underline{coli} and \underline{could} not specifically identify antigen-producing clones. These results indicate that the 65kD protein antigens of \underline{M} . tuberculosis and \underline{M} . leprae are homologues and share a number of epitopes. addition to these shared epitopes, however, both 65kD antigens have epitopes that are specific for one species relative to the other.

No cross-reactivity was observed between other antigens of these two mycobacterial species.

- Because monoclonal antibodies recognize a single epitope and because only one or a few antibodies were available for each antigen, it is not clear whether the 65kD proteins are the only homologous protein antigens of M. tuberculosis and M. leprae.
- Among the antigens for which lambda gtll clones have been isolated, the 18kD antigen of M. leprae and the 19kD antigen of M. tuberculosis are of similar size. To determine whether these two antigens are related, the homology of the DNA sequences that encode these antigens was examined. At conditions of reduces
- antigens was examined. At conditions of moderate stringency, no hybridization was observed between

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the insert DNA and Y3147 (an M. tuberculosis 19kD clone) and Y3179 (an M. leprae 18kD clone). This indicates no significant homology between the DNA sequences of the insert DNAs of these two clones. This result suggests that the M. tuberculosis 19kD and the M. leprae 18kD proteins are unlikely to be homologous.

As a result of the work described, recombinant DNA clones encoding five major protein antigens of M. tuberculosis were isolated through the use of an extensive collection of well-characterized murine monoclonal antibodies. These five proteins were also found to be major antigens in the rabbit humoral immune response to M. tuberculosis. One of these antigens, the 65kD protein, is shared with another other mycobacterial pathogen M. leprae.

Several lines of evidence indicate that the 65kD antigen is among the most immunodominant of the protein antigens of M. tuberculosis. Eleven of the 25 different M. tuberculosis and M. leprae monoclonal antibodies examined in this study recognized the 65kD recombinant antigen from one or both mycobacteria. In addition, almost half of the recombinant DNA clones isolated with rabbit polyclonal anti-M. tuberculosis sera express the 65kD antigen, reflecting the predominance of antibody to this antigen in these sera.

Considerable evidence indicates that the 65kD antigen plays an important role in the human response to tuberculosis. Antibodies directed against this protein can be detected in the serum of

patients with tuberculosis. The 65kD antigen is present in purified protein derivatives (PPD's) of M. tuberculosis, M. bovis, and other mycobacteria. Thole, J.E.R. et al., Infection Immunity, 50:800-806 (1985). Finally, helper T cell clones reactive with recombinant 65kD antigen have been isolated from patients with tuberculosis, indicating that this antigen is involved in the cell-mediated as well as the humoral immune response to tuberculosis.

Dacillus, the 65kD antigens of the leprosy bacillus, the 65kD antigen appears to elicit antibody and T cell responses similar to those observed for the M. tuberculosis antigen. Both serum antibodies and T cells directed against the 65kD M.

- leprae antigen have been observed in patients with leprosy. Britton, W.J. et al., Journal of Immunology, 135:4171-4177 (1985); Mustafa, A.S. et al., Nature, 319:63-66 (1986). In addition, T cell clones from leprosy patients have been found to
- respond to recombinant 65kD protein of M. bovis, as well as to PPD's from both M. bovis BCG and M. leprae. Emmrich, F. et al., Journal of Experimental Medicine, 163:1024-1029 (1986); Shankar, P. et al., Journal of Immunology, 136:4255-4263 (1986). It is
- interesting to note that in vaccine trials in Asia and Africa, BCG provided significant protection against leprosy, ranging from 20% to 80%. Fine, P., Tubercle, 65:137-153 (1984). An intriguing possibility is that the M. bovis BCG 65kD antigen is
- involved in engendering the immune protection

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provided by this vaccine against \underline{M} . leprae, as well as against \underline{M} . tuberculosis.

In addition to the 65kD antigen, there is evidence that the 19kD and 71kD antigens of M. tuberculosis may be particularly important in the immune response to this bacillus. Helper T cell clones from tuberculosis patients have been isolated which respond to the recombinant 19kD protein. The 71kD antigen is recognized by the humoral immune system of both mice and rabbits, and antibody to this antigen has been shown to be a prominent component of hyperimmune anti-M. tuberculosis rabbit sera.

VIII. Isolation of DNA Clones for Genes Encoding Proteins of Additional Mycobacteria

Using the procedures described above for isolation of genes encoding M. tuberculosis proteins, genes encoding proteins of additional mycobacteria were isolated. DNA clones containing genes encoding the following proteins were isolated:

	Mycobacterium	Protein	Clone
	M. bovis BCG	71kD	PL1-101
		65kD	PL1-105
		19kD	PL1-501
25		14kD	PL1-502
	M. smegmatis	65kD	PL1-206
	M. avium	65kD	PL1-401
	M. africanum	65kD	PL1-301

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For purposes of comparison, genes encoding the following proteins were isolated for <u>M. tuberculosis</u> and <u>M. leprae:</u>

	Mycobacterium	Protein	Clone		
05	M. tuberculosis	71kD	Y3272		
		65kD	Y3150		
		19kD	Y3147		
		14kD	Y3248		
	M. leprae	65kD			

10 The following strains were used for this purpose:

	Species	Isolate				
	M. leprae	Armadillo isolate (WHO)				
	M. tuberculosis	Erdmann strain				
15	M. africanum	African clinical isolate				
	M. bovis BCG	Danish vaccine strain				
	M. smegmatis	MC ² -6				
	M. avium	AIDS patient isolate				

DNA from recombinant lambda gt11 clones was isolated, as described above, and mapped with restriction endonucleases, using standard techniques. Davis, R.W. et al., Advanced Bacterial Genetics: A Manual for Genetic Engineering, Cold Spring Harbor (1980).

Figure 4 presents a comparison of the restriction maps for four genes of <u>M. tuberculosis</u> with the restriction maps for four genes of <u>M. bovis</u> BCG which encode proteins of the same molecular weight. As is evident from the figure, in each case, the

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restriction sites on the two genes (e.g., those on the <u>M. tuberculosis</u> gene and those on the <u>M. bovis</u> gene which encodes a protein of the same molecular weight) are essentially identical. This indicates that the sequence of the genes of the two mycobacteria (at least those encoding these four proteins) are very similar and, therefore, the proteins they encode are also very similar.

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Figure 5 presents a comparison of the restriction map for the gene encoding the 65kD protein for the six mycobacteria. As is evident, the restriction maps for the genes encoding the 65kD protein of M. tuberculosis, M. africanum, M. bovis BCG, M. smegmatis and M. avium are essentially identical.

The fact that there is no detectable difference among these mycobacteria at the level of the restriction map is an indication that, at least at this level, the encoded proteins are the same.

As is also evident, the map of the <u>M. leprae</u> 65kD gene has several identical restriction sties in common with those of the other mycobacteria; it also has two sites not found in the other genes and lacks three sites present in the others. This indicates that, at the level of the restriction map, there are similarities in the DNA (and the encoded protein). In addition, however, there are differences apparent at this level.

IX. Diagnostic, Therapeutic and Preventive Applications

The isolation of genes encoding major protein antigens of M. tuberculosis makes it possible to

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address problems which presently exist in diagnosing treating and preventing tuberculosis. Isolation of genes encoding proteins of other mycobacteria, such as M. bovis BCG, M. africanum, M. smegmatis and M. avium makes it possible to address similar problems in diseases which they cause.

The nucleotide sequence of three of the five genes has been determined. The sequence of the remaining genes can be determined using well-known methods, such as that of Sanger et al. Sanger, F. et.al., Proceedings of the National Academy of Sciences, USA, 74:5463-5467 (1977). The amino acid sequence of each of the immunodominant proteins has been deduced from the nucleotide sequence of the three genes and can be done for the others.

Identification and characterization of the genes for major tuberculosis protein antigens and of the proteins themselves make it possible to develop improved reagents for diagnosis and immuno-prophylaxis of tuberculosis. Proteins antigens encoded by an entire gene, or amino acid sequences (e.g., peptides, protein fragments) which make up the antigenic determinant of a M. tuberculosis antigen (i.e., M. tuberculosis-specific antigenic determinants) may be used in serodiagnostic tests and skin tests. Such antigens would be highly specific to the tuberculosis bacillus and the tests in which they are used would also be highly specific. Highly specific serological tests would be of great value in screening populations for

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individuals producing antibodies to M. tuberculosisspecific antigenic determinants; in monitoring the
development of active disease in individuals and in
assessing the efficacy of treatment. As a result,
early diagnosis of tuberculosis will be feasible,
thus making it possible to institute treatment at an
early stage of the disease and, in turn, to reduce
the likelihood it will be transmitted.

As a result of the work described, it is also possible to determine which segment(s) of the M. tuberculosis antigen is recognized by M. tuberculosis-specific T cells. A mixture of peptides recognized by helper T cells can serve as a specific skin test antigen useful in assessing immunological status (delayed hypersensitivity) of infected individuals and those with whom they come in contact. This specific skin test antigen would be useful in evaluating rapidly the immunological efficacy of anti-tuberculosis vaccines.

It is reasonable to expect that the products encoded by M. tuberculosis genes, particularly those shown to be recognized by helper T cells, are themselves immunogenic and thus useful components of vaccines against tuberculosis. These products include proteins and portions of such proteins (e.g., polypeptides and peptides). For example, one approach to vaccine development is the introduction of genes encoding products (e.g., polypeptides) which provide immunological protection into viruses such as vaccinia virus, or bacteria, such as cultivatable mycobacteria, thus producing a vaccine

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capable of engendering long-lasting and very specific immunity. The genes encoding five immunodominant protein antigens of the tuberculosis bacillus, described herein, are useful for that purpose; genes encoding the 65kD, 19kD and 71kD antigens, or a portion thereof, are particularly valuable in vaccine construction.

Because of the similarities in the DNA encoding similarly-sized proteins and, thus, of the encodied proteins themselves, it is possible that, for example, a vaccine effective against two or more of the mycobacteria can be produced.

EXEMPLIFICATION

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Isolation and Analysis of Recombinants Expressing the 65kD M. tuberculosis Antigen

The recombinant DNA library of M. tuberculosis genomic DNA fragments in the lambda gtll vector was constructed as described above. Recombinant phage lambda RY3143 and lambda RY3146 were used. Young, R.A. et al., Proceedings of the National Academy of Sciences, USA, 82:2583-2587 (1985). Subclones of the mycobacterial DNA inserts in these recombinant phage were constructed in pUC19 or M13mp9 vectors using standard recombinant DNA techniques. Messing, J. and J. Viera, Gene, 19:269-276 (1982). Maniatis, T. et al., Molecular Cloning: A Laboratory Manual,

J. and J. Viera, <u>Gene</u>, <u>19</u>:269-276 (1982). Maniatis T. <u>et al.</u>, <u>Molecular Cloning: A Laboratory Manual</u>, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1982).

Monoclonal antibodies specific for the 65kD antigen were obtained from the Immunology of Tuberculosis Scientific Working Group under a grant from the WHO/World Bank/UNDP Special Program for 05 Vaccine Development. These antibodies included IT-13 (WTB-78), IT-31 (SA2D5H4), and IT-33 (MLIIH9). Coates, A.R.M. et al., Lancet, 2:167-169 (1981). Gillis, T.P. and T.M. Buchanon, Immunology, 37:172-178 (1982). Anti-B-galactosidase antibodies 10 were purchased from CooperBiomedical. Polyclonal rabbit antisera directed against a sonicate of M. tuberculosis strain H37Rv were elicited as described by Minden and co-workers. Minden, P. et al., Infect. Immun., 46:519-525 (1984). Results are 15 shown in Table 2.

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TABLE 2: PATTERNS OF ANTIBODY REACTIVITIES

	Number of Clones	Reactivity with Antibodies			
	•	<u>IT-13</u>	IT-31	IT-33	
	27	+	+	+	
05	1	+	+	+	
	2	+	-	+	
	3	-	+	+	
	1	+	-	_	
	2	-	+	· 	
10	2	-		+	

a: Recombinant clones expressing antigens reactive with the 65kD antigen specific monoclonal antibodies IT-13, IT-31, and IT-33 were isolated as described above. For the initial screen, a pool of the three antibodies was used; it contained a 1:1000 dilution 15 of each antibody to screen a total of about 8×10^5 recombinant phage from the lambda gtll-M. tuberculosis library. To determine which monoclonal antibody reacted with which of the 38 plaque-20 purified recombinants, about 100 pfu of each recombinant phage were inoculated in small spots on a lawn of Y1090. The phage were allowed to grow and induced to synthesize the foreign proteins as described previously. The filters were then reacted 25 with a 1:1000 dilution of one of the monoclonal hybridoma antibodies as described above.

The lambda gtll-M. tuberculosis library was screened with the monoclonal antibodies specific for the 65kD antigen and clones reactive with them were isolated essentially as described by Young et al. Young, R.A. et al., Proceedings of the National 05 Academy of Sciences, USA, 82:2583-2587 (1985). Briefly, for each 150mm LB plate, 0.6ml of a fresh overnight culture of Y1090 was infected with 1-2 x10⁵ plaque forming units of the library. After 3.5-4 hours of growth at 42°C, the plaques were 10 overlaid with a dry nitrocellulose filter which had been saturated with 10mM isopropyl-B-D-thiogalactopyranoside (IPTG). The plates were incubated an additional 3.5-4 hours at 37°C and then removed to room temperature and the position of the filters 15 marked. The filters were washed briefly in TBST (50 mM Tris-HCl, pH 8, 150mM NaCl, 0.05% Tween 20) and then incubated in TBST + 20% fetal calf serum. After 30 minutes at room temperature, the filters were transferred to TBST plus antibody. For the 20 initial screen, the antibody mix contained a 1:1000 dilution of IT-13, IT-31, and IT-33. The filters were incubated with the antibody solution overnight at 4°C with gentle agitation, washed in TBST and reacted with biotinylated goat anti-mouse immuno-25 globulin, the Vectastain ABC reagent, and developer as described by the manufacturer (Vector Laboratories). After the color had developed the filters were washed with several changes of water and air dried. Phage corresponding to positive 30 signals were twice plaque purified. To determine

which monoclonal antibodies reacted with which of the recombinant phage, about 100 pfu of a purified phage stock were inoculated in a small spot on a lawn of Y1090 bacteria on an LB plate. The phage were allowed to grow and induced to synthesize the foreign proteins as described above. The filters were then reacted with a 1:1000 dilution of one of the monoclonal antibodies. The subsequent steps were the same as for the initial screen.

10 Western blot assays were carried out as follows: Cells containing phage or plasmids in which the expression of the foreign sequences was under the control of the E. coli lac gene regulatory sequences were induced to synthesize the foreign 15 proteins by incubating the cells in the presence of 2.5mM IPTG for 2 hours. Crude lysates of cells expressing lambda gtll recombinants were made as described in Huynh et al. Huynh, T.V. et al., In: DNA Cloning Techniques: A Practical Approach, (D. 20 Glover, ed.) IRL Press, Oxford, Vol. 1, pp. 49-78 (1985). Crude lysates of cells expressing plasmid encoded proteins were made by harvesting cells from overnight cultures and resuspending the cells in 10 mM Tris pH7.5/10 mM EDTA containing 100 ug 25 lysozyme/ml. After 10 minutes at room temperature, SDS was added to a final concentration of 0.5%.

lysozyme/ml. After 10 minutes at room temperature, SDS was added to a final concentration of 0.5%. A protease inhibitor (Trasylol, Boehringer Mannheim) was added to all crude lysates at a final concentration of 0.3%. The crude protein preparations were electrophoresed on 10% polyacrylamide-SDS

were electrophoresed on 10% polyacrylamide-SDS

Laemmli gels and the separate proteins electrophor-

etically transferred to nitrocellulose. Laemmli,
U.K., Nature, 227:680-685 (1970). Towbin, H. et

al., Proceedings of the National Academy of

Sciences, USA, 76:4350-4354 (1979). The immobilized

proteins were reacted with a 1:1000 dilution of

monoclonal antibody IT-13 in TBST overnight at 4°C.

The nitrocellulose filters were then washed, reacted

with peroxidase-conjugated goat anti-mouse immuno
globulin, and developed as described by Niman and

co-workers. Niman, H.L. et al., Proceedings of the

National Academy of Sciences, USA, 80:4949-4953

(1983).

The sequences of 5'-end-labeled restriction fragments of the mycobacterial DNA were determined 15 by a modification of the partial chemical degradation technique of Maxam and Gilbert. Brow, M.A.D. et al., Mol. Biol. Evol., 2:1-12 (1985). Maxam, A.M. and W. Gilbert, Proceedings of the National Academy of Sciences, USA, 74:560-564 20 (1976). For the M13/dideoxy sequencing studies, Sau3AI fragments from the mycobacterial DNA inserts were subcloned into the BamHI site of Ml3mp9. Phage DNA was isolated from the M13 recombinants and subjected to the dideoxy chain termination sequencing reactions. Biggin, M.D. et al., 25 Proceedings of the National Academy of Sciences, USA, 80:3963-3965 (1983). Sanger, F. et al., Journal of Molecular Biology, 143:161-178 (1980). The products of the sequencing reactions were

electrophoresed on 6% acrylamide/7M urea/0.5-2.5 x TBE gradient sequencing gels. The gels were dried

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under vacuum and exposed to Kodak XRP-1 film. nucleotide sequences were determined independently for both strands of the mycobacterial DNA.

Computer-aided analyses of the nucleic acid sequences and deduced protein sequences were performed using the Databases and programs provided by the Nucleic Acid and Protein Identification Resources of the National Institutes of Health as well as the programs of Chou and Fasman and Hopp and 10 Woods. Chou, P.Y. and G.D. Fasman, Adv. Enzym., 47:45-148 (1978). Hopp, T.P. and K.P. Woods, Proceedings of the National Academy of Sciences, USA, 78:3824-3828 (1981). The nucleotide sequence of the region containing the M. tuberculosis 65kD gene and the deduced amino acid sequence of the two long open reading frames are represented in Figure ε.

B-galactosidase assays were also carried out. Cells were grown in LB broth or LB broth plus 2.5mM IPTG to an OD of about 0.3. Crude lysates were made and b-galactosidase activity assayed as described by Miller. Miller, J.H., Experiments in Molecular Genetics, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1972).

25 Equivalents

Those skilled in the art will recognize or be able to ascertain, using no more than routine experimentation, many equivalents to the specific materials and components described herein. equivalents are intended to be encompassed in the scope of the following claims.

CLAIMS

- 1. Isolated DNA encoding an immunogenic protein antigen of Mycobacterium tuberculosis.
- 2. DNA of Claim 1 selected from the group consisting of DNA encoding Mycobacterium tuberculosis protein antigens of molecular weight 71kD, 65kD, 19kD, 14kD and 12kD.
- 3. Isolated DNA encoding an immunodominant protein antigen of Mycobacterium tuberculosis, the protein antigen having a molecular weight of approximately 65kD and recognized by a monoclonal antibody selected from the group consisting of: IT-31; C1.1; IIH9; IIC8; T2.3; Y1-2; SA2.D7C and IT-13.
- 15 4. Isolated DNA encoding an immunodominant protein antigen of Mycobacterium tuberculosis, the protein antigen having a molecular weight of approximately 19kD and recognized by a monoclonal antibody selected from the group consisting of: IT-10; IT-12; IT-16; and IT-19.
 - 5. Isolated DNA encoding an immunodominant protein antigen of Mycobacterium tuberculosis, the protein antigen having a molecular weight of approximately 71kD and recognized by the monoclonal antibody IT-11.

- 6. Isolated DNA encoding an antigenic determinant of Mycobacterium tuberculosis protein.
- 7. DNA of Claim 6 which encodes an antigenic determinant selected from the group consisting of antigenic determinants of Mycobacterium tuberculosis proteins of molecular weight 71kD, 65kD, 19kD, 14kD and 12kD.
- 8. Isolated DNA encoding an amino acid sequence of an antigenic determinant of Mycobacterium
 10 tuberculosis protein, said protein having a molecular weight of approximately 65kD.
 - 9. Isolated Mycobacterium tuberculosis DNA encoding an immunodominant protein antigen having a molecular weight of approximately 65kD, said DNA selected from the group consisting of:
 - a. the DNA insert of clone Y3141;
 - b. the DNA insert of clone Y3143:
 - c. the DNA insert of clone Y3150;
 - d. the DNA insert of clone Y3253; and
- e. the DNA insert of clone Y3262.
 - 10. A protein antigen encoded by DNA of Claim 9.
 - 11. A protein antigen of Claim 10, wherein the protein antigen is recognized by a monoclonal antibody selected from the group consisting of IT-31; C1.1; IIH9; IIC8; T2.3; Y1-2; SA2.D7C and IT-13.

- 12. Isolated DNA having a nucleotide sequence selected from the group consisting of: a) the nucleotide sequence represented in Figure 6, or a portion thereof; b) the nucleotide sequence represented in Figure 7, or a portion thereof; and c) the nucleotide sequence represented in Figure 8, or a portion thereof.
- 13. A protein or a peptide selected from the group consisting of: a) proteins or peptides encoded by the nucleotide sequence represented in Figure 6, or a portion thereof; b) proteins or peptides encoded by the nucleotide sequence represented in Figure 7, or a portion thereof; and c) proteins or peptides encoded by the nucleotide sequence represented in Figure 8, or a portion thereof.
 - 14. A peptide having the amino acid sequence of an antigenic determinant of Mycobacterium tuberculosis protein, said antigenic determinant being unique to Mycobacterium tuberculosis protein.
 - 15. A peptide encoded by isolated Mycobacterium tuberculosis DNA, said peptide recognized by helper T cells.
- 25 16. A peptide encoded by the <u>Mycobacterium</u> tuberculosis DNA insert of clone Y3150 or a portion of said DNA insert.

- 17. Isolated DNA encoding a protein of Myco-bacterium africanum the protein having a molecular weight of 65kD.
- 18. Isolated DNA encoding a protein of Myco-bacterium avium, the protein having a molecular weight of 65kD.
 - 19. A vaccine comprising DNA encoding Mycobacterium tuberculosis protein in a recombinant vaccine vector capable of expressing said DNA.
- 10 20. A vaccine of Claim 19 in which the recombinant vaccine vector is vaccinia virus or cultivatable mycobacteria.
- 21. A vaccine of Claim 20 in which the DNA encodes the 65kD Mycobacterium tuberculosis protein recognized by the monoclonal antibody IT-13, or a portion of said protein.
 - 22. A vaccine comprising DNA encoding an antigenic determinant unique to Mycobacterium tubercu-losis cultivatable mycobacteria capable of expressing said DNA.
 - 23. A method of detecting antibody against Myco-bacterium tuberculosis in a biological fluid, comprising the steps of:
- a) incubating an immunoadsorbent comprising a solid phase to which is attached

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immunodeterminant Mycobacterium tuberculosis protein with a sample of the biological fluid to be tested, under conditions which allow the anti-Mycobacterium tuberculosis antibody in the sample to bind to the immunoadsorbent;

- b) separating the immunoadsorbent from the sample; and
- c) determining if antibody is bound to the immunoadsorbent, as an indication of anti-Mycobacterium tuberculosis in the sample.
- 24. A method of Claim 23 in which the Mycobacterium tuberculosis protein attached to the solid phase has a molecular weight of approximately 65kD.
- 25. A method of detecting antibody against Mycobacterium tuberculosis in a biological fluid, comprising the steps of:
 - a) incubating an immunoadsorbent comprising a solid phase to which is attached a peptide having the amino acid sequence of an antigenic determinant of Mycobacterium tuberculosis protein with a sample of the biological fluid to be tested, under conditions which allow antibody against Mycobacterium tuberculosis to bind to the immunoadsorbent;
 - b) separating the immunoadsorbent; and
 - c) determining if antibody is bound to the immunoadsorbent, as an indication of the

presence of the antibody against Mycobacterium tuberculosis in the sample.

- 26. A method of Claim 25 in which the peptide has the amino acid sequence of an antigenic determinant which is unique to Mycobacterium tuberculosis protein.
- 27. A kit useful in detecting antibody against

 Mycobacterium tuberculosis in a biological

 fluid, comprising a collection of reagents for

 immunoassay of said antibody, said collection

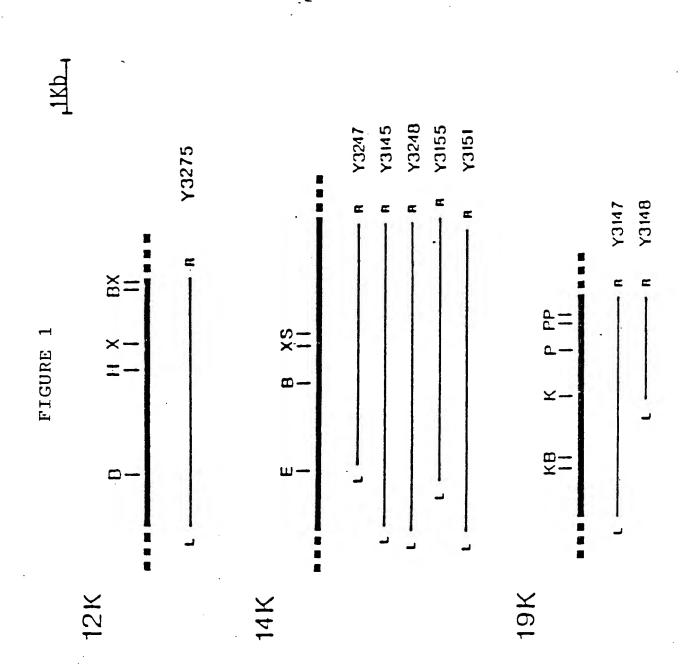
 of reagents a solid phase to which is attached

 immunodeterminant Mycobacterium tuberculosis

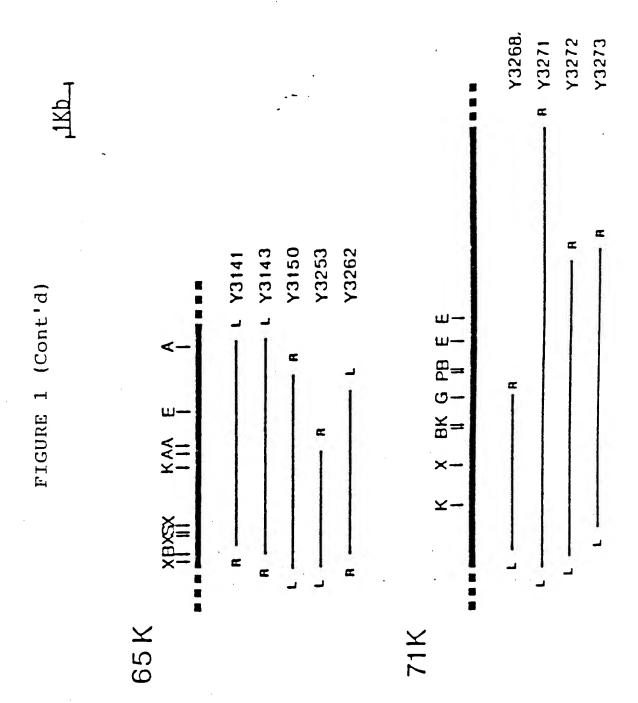
 protein or a peptide having the amino acid

 sequence of an antigenic determinant of

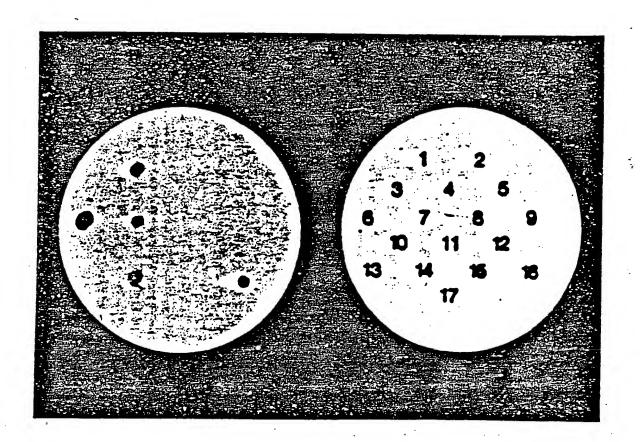
 Mycobacterium tuberculosis.



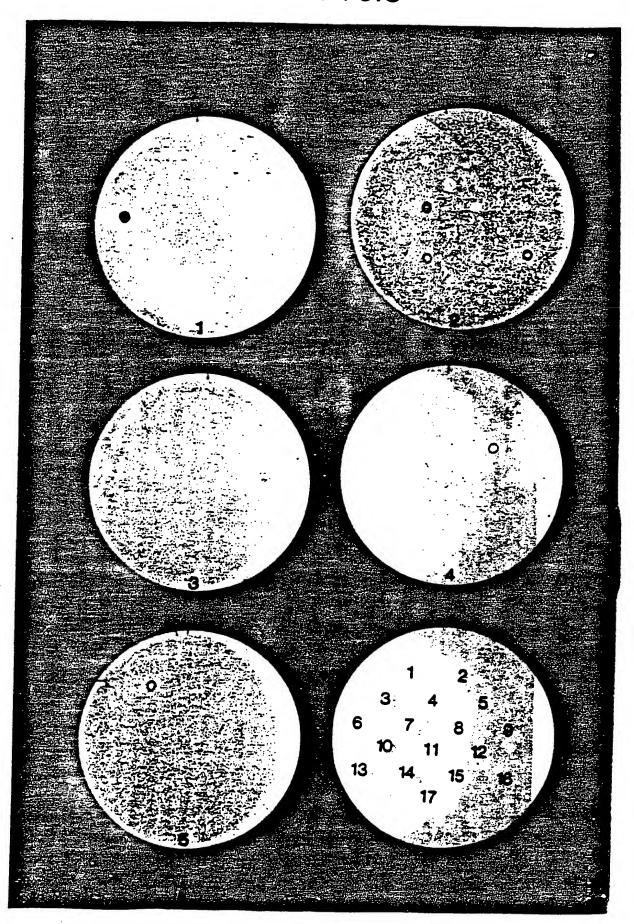


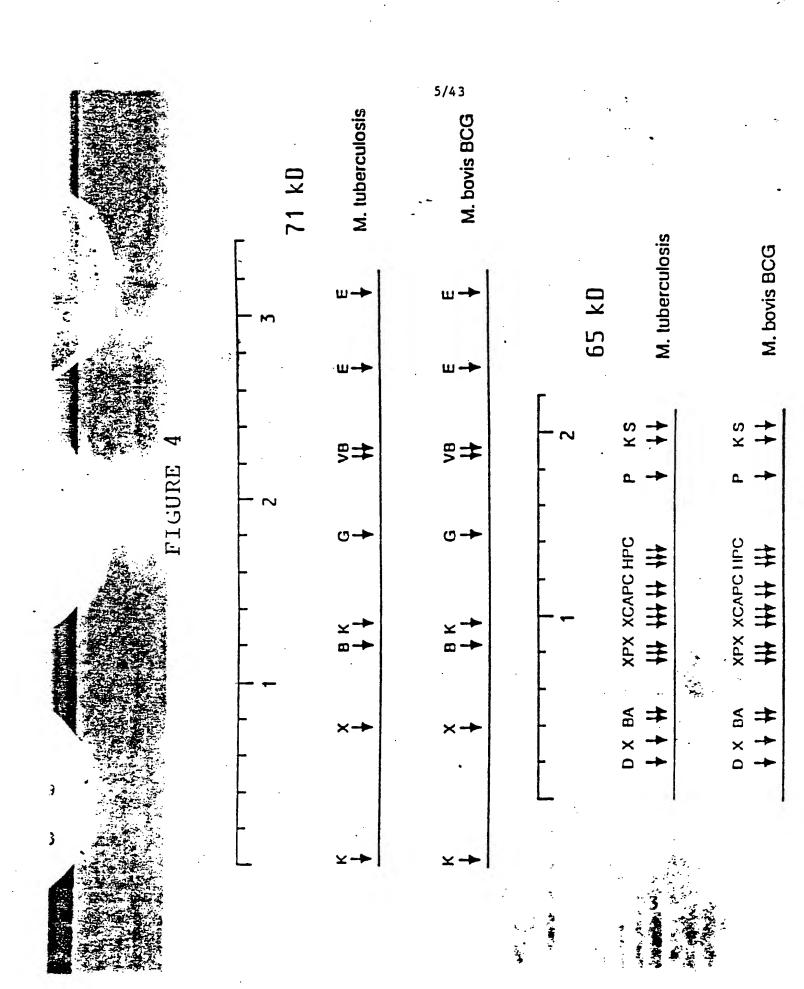


3/43 FIG.2

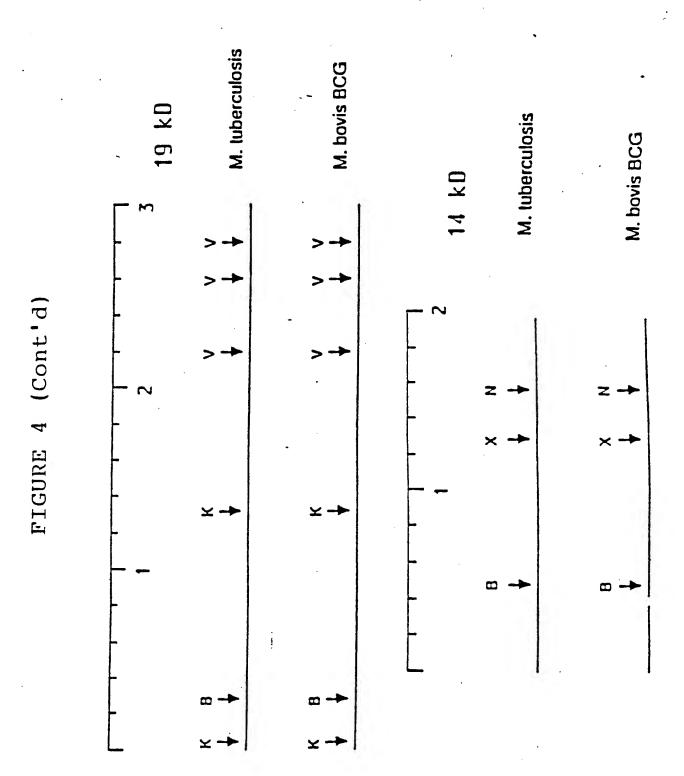


4/43 FIG.3

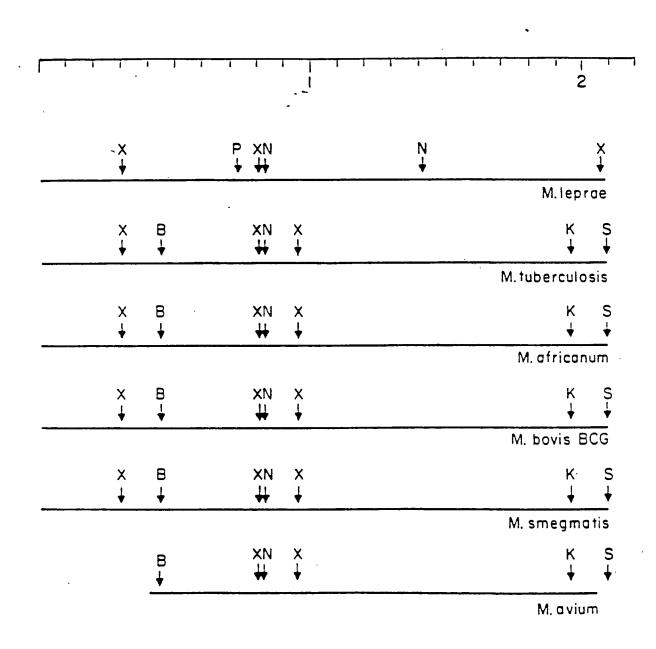








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65 kD Gene

FIG. 5

FIGURE

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9 TICCGIGICAAICGCCICGGIGAGAIAGCCCGACCAGGCGCGCGGGAICGCGCACCAAGGC 50 只 Ω K G 3 S 民 G S 回 20 K 回 Ø Ω ہنا

CGGCGCGTCCCAGTCGTCCAACACGTCCAATACGCACCGCAAAAGCCGGTACGTGTTGCG 110 Σ Ы 100 Σ 90 80 Z Q a 70 Ø K G

CGCCTTCGCGCCCTTTATCTTCACGTAGAGGCGTCCCCGGAGCCACGGGCCCCCGGGTAGAC Σ U 170 U H ū GCGGAAGCGCGGGAAATAGAAGTGCATCTCCGCAGGGGCCT 160 K U ď K K 150 ы Σ Ξ S 140 بتا ᄓ Ŀı ы K 130 K ū U K

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240 CTTGAGCTAGAGGTGGGGCAGCTACCACACCAGAGGGGGCCAGAGCCACTACAGCTGGCA 230 U 回 GAACTCGATCTCCACCCGTCGATGGTGTGGGTCTCCCCGGTCT G ĿJ r S 200 C U U > 190 只 П ы ы لتا

FIGURE 6 (CONT'D)

10/43 300 GGGGGCCTCTGGCGCGCGGTTGTGCCACGGCATGTACATCGGGCGTGCCGCGTAGTAGTAGCG CCCCCGGAGACCGCGCGCCAACACGGTGCCGTACATGTAGCCCGCACGGCGCATCATCGC CGGCACGACGAGCGAAACCTCACCGGTCGACAGTGTCTGCCCGAGGCCGCAGCCGACGTG GCCGTGCTGCTCGCTTTGGAGTGGCCAGCTGTCACAGACGGGCTCCGGCGTCGGCTGCAC Σ Σ ĸ 350 290 U K [L] ۵ U U ഗ 340 G 280 Ø U Σ Ø 330 270 U ഗ U H 320 260 Н u < Z > ĸ ĸ G u 310 250 GR Д 民 Д Д

GCTCGGCCGCATCTACAAAAGGACGTGCCGCACGCGCCACTTGGGGAGGCCGCGGTCGTG CGAGCCGGCGTAGATGTTTCCTGCACGGCGTGCGCGGTGAACCCCTCCGGCGCCAGCAC K Ή U Ы ū 400 U 390 Ø 380 Ы Ω U ഗ

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GCGGTGGAAAGGGCGCAGGTGCAGCCGGACCCACTGCGGGTTCGTGGGGTGGCTTTAC CGCCACCTTTCCCGCGTCCACGTCGGCCTGGGTGACGCCGAGCACCCCACCGAAATG U 470 Н U 460 Q ¥ K Ω Ω K U U ¥

540 TAGCTGTACCGACACCCACATCTACTGGCGCTGGTGCCCCGCCAGCCGAGGCGCCACCCG G. R 530 S ¥ K Ω 520 U G Д 3 510 G 500 G Q S 490 Σ

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009 CGCTATGTTCAGGTCGCGCCGCCGGTGGAGCCACCTGTGGTTGCCCAGCTACTGCTA 590 а GCGATACAAGTCCAGCGCGGCGGCGCCACCTCGGTGGACAC 580 S ഥ 570 C K 560 K K K Ч C Ω 550 K

GTCGGGTCACAGTGGGAGTTGCTTCGACTATAACCTCTATAGCTTAGGCGCCTGGGACTAT CAGCCCAGTGTCACCCTCAACGAAGCTGATATTGGAGATATCGAATCCGCGGGACCTGATA Ω 650 M 640 G Z 630 ഗ ᄓ Z ٠ ت U ŋ U

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GATGCCCGGCACCACCTGGTAGAGGCCCTGTTTCGCGGTCAGCTGGGATTGCCGCCACAG CTACGGGCCGTGGTGGACCATCTCCGGGACAAAGCGCCAGTCGACCCTAACGGCGGTGTC 710 Ø S Q ľ Q 700 K × 069 <u>ෆ</u> u 089 019 U

14/43 780 CGACCCTACGTGGCTACAGCCGCGCGTGGCAGCTCTTTGCTCATGCGCAGCAACAGGGT 3 U Ω \mathcal{O} Z GCTGGGATGCACCGATGTCGGCGCGCGCGTCGAGAAACGAGTACGCGTCG1 G 770 Z Ы > ဟ S 760 Д J ū 民 Ω Z 750 U Н K K U K K S K 740 Ы H H Σ 区 ဟ 3 工 U 730 Σ 区 K 3 S

TGATCACACGGGGACAGCGCGGCAATGAATCCGCG GTGGTGCGCTGGTAGCCGTCGGAACTAGTGTGTGCCCCTGTCGCGCCGTTACTTAGGCGC G 回 830 4 K 820 U 出 Д 810 Ω 800 U K CACCACGCGACCAT G 790 又 3 > 工

900 TAGCCGCAGCAGCTTTAGGCAACACAGTACGTTGCCATTGCTCACAAGTGGCACACGGCG TCACCGI 890 U ATCGGCGTCGTCGAAATCCGTTGTGTCATGCAACGGTAACGAGT 880 a 870 Σ Σ Ø 860 ы Ŀı 850 ¥

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TGCCACTACTACGGTG GACCTACTGCCGTCACCCTCCAAACACAAGGTAGCCGTGATGTAACGGTGATGATGCCAC 950 G Z GTTCCATCGGCACTACAT 930 回 CTGGATGACGGCAGTGGGAGGTTTGT 920 S 910 Σ Ø

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FIGURE 6 (CONT'D)

1020 1080 GTGCGGCCATCTACGGCAACCGCTTGGTGCGATGGCTGGTCTTTCTCTCTTAAAAGGCGG GCACCTAGACCTCGGGCCCTGCTAACGCGCATACTGCCGAAGCGGTCCTCAATGCCGATG U U 回 1010 1070 K ဟ 1060 1000 3 Ü Н K CACGCCGGTAGATGCCGTTGGCGAACCACGC 1050 990 U S C 1040 3 980 Д S 970 1030 U

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6 (CONT'D)

FIGURE

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CTGGCGATGCTGTCCGTTTCCTCGTGTCCCACTTCGCACCTGACTGCCAGCGCCATCGGC a S K Q GACCGCTACGACAGGCAAAGGAGCACAGGGTGAAGCGTGGACT S 1120 S ഗ S 1110 \mathcal{O} U 4 S 1100 凶 U Д K 3 Ŀ Ω 1090 U Ω F

1200 TCAAGCAACAAGTCGACTACAGGAA CICGGCGGTAAGACCAGCGICCAGAAAGGCCIACAAGIICGIIGIICAGCIGAIGICCII 1190 S Ö 1180 Z CTTTCCGGATGT C Д U E1 × K GAGCCGCCATICIGGICGCAGGI Ω U Д 1160 ď H Д O ы 1150 Z Σ 3 ĸ П

Д 1250 ڻ ا 1230 K ഗ 1220 S 1210 G

18/43

AGGTCGTCATCGACGGTAAGGACCAGAACGTCACCGGCTCCGTGGTGTGCACAACCGGG TCCAGCAGTAGCTGCCATTCCTGGTCTTGCAGTGGCCGAGGCACCACACGTGTTGGCGCC 1310 1300 တ K 1290 G K ĸ 1270 只

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FIGURE 6 (CONT'D)

ACGGCAACCCTCCGGAGGTGAAGTCCGTTGGGCTCGGTAACGTCAACGGCGTCACGCTGG TGCCGTTGGGAGGCCTCCACTTCAGGCAACCCGAGCCATTGCAGTTGCCGCAGTGCGACC GGCCGTTACAGTTGTAGCGCTAGCCGCCCCGCCGCTGGCCGTAACGGCGGCACGAGTGGC Ø 1430 1370 民 U K 1420 1360 K K G Д 1410 1350 C G Ø K S ഗ 1400 1340 S K S Σ U ഗ 1390 1330 а

19/43

1500 CTATGTGCAGCCCGTGGCCTGTCCCATTGCGGAGCCGTTGGTTCCTGCCGTCGGTGATGT GATACACGTCGGGCACCGGACAGGGTAACGCCTCGGCAACCAAGGACGGCAGCCACTACA . ა 1490 K 1480 K 1470 O 1460 G K 1450 H Ω

20/43

1560 TCTAGTGACCCTGGCGATGGCCCCAGCTGTACCGGTTGGGCTACAGTGGCCACTTGTTCA AGATCACTGGGACCGCTACCGGGGTCGACATGGCCAACCCGGTGTCACCGGTGAACAAGT S 1550 1540 3 3 1530 U م 1520 K ഗ 1510

21/43 GCAAGCTTTAGCTCCACTGGACAAGGATTGGATTTCGCACAGCTACGCCCGACACTTGTC CGTTCGAAATCGAGGTGACCTGTTCCTAACCTAAAGCGTGTCGATGCGGGCTGTGAACAG S Q 1610 Ľ Σ Ω S > 1600 × 1590 Z О U H Ø 只 1580 ഗ K 只 1570 × S ы

GCGCAGCCTCGGCCCGTCAGTCCGGATCGCGCCGCTGCTAAGCTCGCCAACGGTAGGCAG CGCGTCGGAGCCGGGCAGTCAGGCCTAGCGCGGCGACGATTCGAGCGGTTGCCATCCGTC U 3 Ø 1670 民 Ч 回 ഥ 1660 ഗ S Ы 1650 G G 1640 口 K Ч K G 1630 ഗ U Ω V

TTCACCGTTGGCGTGGCGTTTGAGCCATATAGGCCCACTCGATGAGTGCCACTAGCAAGG AAGTGGCAACCGCACCGCAAACTCGGTATATCCGGGTGAGCTACTCACGGTGATCGTTCC 1730 ഗ C U ы 1700 υ 1690 r 3 М

22/43

GTTGTGCGCCTTGACCACAGCGGAGACGATCGCCAGGCCGGAGCCCGGTGCTACCGGCTTG CAACACGCGGAACTGGTGTCGCCTCTGCTAGCGGTCCGGGCTCGGGCCACGATGGCCGAAC 1790 ГJ 1780 Ω U S 1760 Ω 1750 K Z

GCCTTATGGGCCGGGTAACAGCTAGTGGACGTCGTGCTGCACGCAGCCGGGCCACGAGTT

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FIGURE 6 (CONT'D)

ĸ 1860 CGCCCTGGCACTGCATAGCGGCGCCCGCTTGGCGAGCTTTTGGAGCCTGACGTCGCGCCG U CGGAATACCCGGCCCATTGTCGATCACCTGCAGCACGACGTGCGTCGGCCCGGTGCTCAA CGGACTGCAGCGCGGC ø 1850 1910 S G ы CGANAACC U 1900 1840 M K GCGGGACCGTGACGTATCGCCGCGGGGGGAACCGCT ď Q 1890 1830 u S K K ہم G 1820 1880 K Ω H S 1870 1810 K S C G α 只

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GTGGTGGACCACGTTGGCACGGCGTAGTGGGCCCTACTGGTGGCCAAGCCCCCCGTCCAG CGCGCAGCAGTGCTAGCACGGCCCTGGCCACACGTGCGCCCGCAACCGGTCGTCCAACCA CACCACCTGGTGCAACCGTGCCGCATCACCGGGATGACCACCGGTTCGGGGGGGAGGTC Z Ļ U U 1970 2030 U ы Q Z K GCGCGTCGTCACGATCGTGCCGGGACCGGTGTGCACGCGGGCGT 1960 2020 区 Ω > Ξ \simeq 2010 1950 G U S 1940 C 2000 U Д K S Ø Ω 1930 1990 Ø Ξ

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(CONT,D)

FIGURE

25/43

CTCGCGGGTGACCACTAGACCAGGCCGTTGTACTCGCAGGCGACTGGCGCAGTTGGCGTTC GAGCGCCCACTGGTGATCTGGTCCGGCAACATGAGCGTCGCTGACCGCGTCAA 2090 K Q S 2080 S Σ 工 2070 Z C G S 3 2060 Ω S Ξ ىم Q 2050 S 3 K K П ĿJ

TCCAGCGGCCGCCCGAGTCCAGACGCGCCAGCAG Ø Ø CCGCGACATGTCCACCGGTCCGCGT 田 2

2160 GGCGCTGTACAGGTGGCCAGGCGCAAGGTCGCCGGGGGGGCTCAGGTCTGCGGGGGTCGTC 2150 ĸ S 2140 2130 ы \simeq 2120 G 5 2110 Σ S

26/43 **E**⊣ 2280 GTCGTCCAGGAGCTGCTCTGCACAATAGGCGAGCCAGAGGCTACGGTGGGCCGAGTAGCG CACACGCTGCCGGAGCCCTAGCAGGGGATAGGCGACGCAGTTAAGGCACATTGGTGCCTA TATCCGCTCGGTCTCCGATGCCACCCGGCTCATCGC K Σ Ŋ, C ы K 2200 2260 ы H GIGIGCGACGGCCICGGGAICGICCCCTAICCG ĸ **⊡** 2190 2250 以 U CAGCAGGTCCTCGACGAGACGTG1 Ω K 2240 2180 S ы K 2230 Ω H

G (CONT'D

FIGURE

27/43

GCGGCGTTCCCCTCAAGCGTCAAGTACTGACCGTAGCCGTTGCTTGACCGCGTGTGCCCA CGCCGCAAGGGGAGTTCGCAGTTCATGACTGGCATCGGCAACGAACTGGCGCACACGGGT K 2330 Ø Ŀ Z 2320 2310 S Σ ы Ø 2300 凡 S ഗ م U 2290 H

2400 AAGTGACCGGACGGCGCGCTGTCACGCCGTCGCTACACCAGCTCGTAGGCCAACTCGCG TTCACTGGCCTGCCGCCGACAGTGCGGCAGCGATGTGGTCGAGCATCCGGTTGAGCGC Z 2390 Σ Ы 2380 Ω ഗ 2370 U Ø K 2360 ഗ K Ø 2350 Y S ſτι

GCTGGGCTCAACGGGGTGGAGCCACCTCCCCAAACGCAGTCCAAGCCCGTGGGCCTGGCC 2460 TCGGGCACCCGGACCGG 2450 U CGACCCGAGTTGCCCCACCTCGGTGGAGGGGTTTGCGTCAGGT 2440 K Ω Z 2430 C G Ы S 2420 ঢা C G Ø 2410 G Ω S

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2520 2510 回 ഗ ы 2500 S Ω. 2490 3 2480 U 只 ū ×

29/43 **AACGCAATCACCGTGACGATTCCGAAAATGATCAGCATCTGCAACATCGTGGCGTCGACG** TTGCGTTAGTGGCACTGCTAAGGCTTTTACTAGTCGTAGACGTTGTAGCACCGCAGCTGC K 2630 Ξ 2620 K 工 K Z 2600 K H G 2590

ы K Ξ а

(CONT'D 9 FIGURE

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K

CGCCGCAACGGCGCCAGCGCCCGCTTGATGACGATTCCGGCGGGGTCGTCGCGGCGACC

GCGGCGTTGCCGCGGTCGCGGGCGAACTACTGCTAAGGCCGCCGCCAGCAGCGCCGCTGG

K U U

FIGURE 7

			3	0/43				
48		96					192	
GAG	Glu	ATC	Ile	ATT CCG CAG ATC GAG GTC ACT TTC GAC ATC 144	Ile		GGC 192	Gly
CGT	Arg	ວອອ	Gly	GAC	Asp		ACC	Thr
GAG	Glu	ACC	Thr	TTC	Phe		၁၅၅	G1y
CAG ATC CAG GTC TAT CAG GGG GAG CGT	Gln Ile Gln Val Tyr Gln Gly Glu Arg	GAG CTG ACC GGC	Leu Leu Gly Ser Phe Glu Leu Thr Gly	ACT	Ile Pro Gln Ile Glu Val Thr Phe Asp	•	GTC ACC GCC AAG GAC AAG GGC ACC	His Val Thr Ala Lys Asp Lys Gly Thr
CAG	Gln	GAG	Glu	GTC	Val		GAC	Asp
TAT	Tyr	TTC	Phe	GAG	Glu		AAG	Lys
\mathtt{GLC}	Val	CTC GGG TCC TTC	Ser	ATC	I1e		229	Ala
CAG	Gln	555	G1y	CAG	Gln		ACC	Thr
ATC	Ile	CTC	Leu	ງລວ	Pro		GTC	Val
CAG	Gln	TTG	Leu		I1e		CAC	His
GTG	Val	AAG	Lys	999	$_{\rm G1y}$		\mathtt{GTG}	Val
TTC CAA CCG TCG GTG	Pro Ser Val	CAC AAC AAG	Ile Ala Ala His Asn Lys	ງງວ	Pro Ala Pro Arg Gly			Asp Ala Asn Gly Ile Val
ອວວ	Pro	CAC	His	ວວວ	Pro		ລອອ	G1y
CAA	Phe Gln	525	Ala	ອວອ	Ala		AAC	Asn
${ m TTC}$	Phe	225	Ala	500			229	Ala
1 GAA	Glu	49 ATC GCC GCG	Ile	97 CCG CCG GCG	Pro		145 GAC GCC AAC GGC ATT	Asp
-		49		97			145	

240		. 288	,	., - J	336			384	
GAA	Glu	GAT	Asp		TTG	Leu	·.	GGT	Gly
CGA ATC CAG GAA GGC TCG GGC CTG TCC AAG GAA	Ile Arg Ile Gln Glu Gly Ser Gly Leu Ser Lys Glu	ATC AAG GAC GCC GAA GCG CAC GCC GAG GAG	Ile Asp Arg MET Ile Lys Asp Ala Glu Ala His Ala Glu Glu Asp		GCC GAT GTT CGT AAT CAA GCC GAG ACA TTG	Ala Asp Val Arg Asn Gln Ala Glu Thr Leu		GAG	Phe Val Lys Glu Gln Arg Glu Ala Glu Gly
$\mathbf{I}^{\mathbf{C}}$	Ser	GAG	Glu		GAG	Glu		CCC	Ala
·CTG	Leu	၁၁၅	Ala		CCC	Ala		TTC GTC AAA GAA CAG CGT GAG GCC GAG	Glu
ეეე	G1y	CAC	His		CAA	Gln		CGT	Arg
\mathbf{TCG}	Ser	gcg	Ala		AAT	Asn		CAG	Gln
ეეე	${\tt Gly}$	GAA	Glu		CGT	Arg		GAA	Glu
GAA	Glu	ວວອ	Ala		ЦLЫ	Val		AAA	Lys
CAG	Gln	GAC	Asp		GAT	Asp		GTC	Val
ATC	Ile	AAG	Lys		CCC	Ala		TTC	Phe
CGA	Arg	ATC	Ile		GAG	Glu			Lys
ATC	Ile	ATG	MET			Arg Lys Arg Arg Glu Glu		CAG ACG GAG AAG	Thr Glu
GAG AAC ACG ATC	Lys Glu Asn Thr	GAC CGC ATG	Arg		CGC GAG	Arg		ACG	Thr
AAC	Asn	GAC	Asp		CGT	Arg			Gln
GAG	Glu	GAC ATT	Ile		CGC AAG	Lys	-	TAC	Tyr
AAG	Lys	GAC	Asp		၁၅၁	Arg		GTC	Val
33		Ţ			6			. 7	

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FIGURE 7 (CONT'D)

432		
\mathtt{GTG}	Val	
SCG	Ala	
CCC	Ala	
GAT	Asp	
GTT	Val	
AAG	Lys	
AAC	Asn	
CTG	Leu	
ACG	Thr	
GAC	Asp	
GAA	Glu	
CCT	Pro	
GTA	Val	
AAG	Lys	
TCG	Ser	
GGT	$_{ m G1Y}$	
382		

480	
CAT CAA	His Gln
CCC C	e Gly H
TTC	Phe
TAT	$\mathbf{T}\mathbf{Y}\mathbf{r}$
GGA	G1y
CGG ATC	Ile
SSO	Trp Arg
TGG	
ACT	Thr
299	$_{ m G1y}$
299	G1y
GAA	Glu
SCG	Ala
GAA	Glu
SSS	Ala
133	

32/4	528 528	
	AGC	Ser
	GCA	
. - I-	999	Glv Ala
•	\mathtt{TCT}	Ser
l	ეენ	Gly
ı	GCA	Ala
	GTC	Val
	GGA	Gly
	CCA	Pro
	999	G1y
	GCT	Ala
	GAA	Glu
	GGA	$_{ m G1y}$
	GAT	Asp
	CCC	${ m G1y}$
	GTC	Val
	181	

	•
576	
CCA	Pro
TGC	Cys
ລອລ	Arg Cys
TGG	Trp
CAC	Gly His Trp
299	Gly
ACA	Thr
\mathtt{GTC}	Val
\mathtt{TGC}	Cys
255	G1y (
TCA	Ser
AGC	Ser
AGC	Ser
CGA	Arg
CTA	Leu
GAT	Asp
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ეყე Leu Gly CGG CTC Pro Arg CCA CCC Pro TGCCys CGG Arg GGG G1y229Ala Arg CGG CGG CGA Arg Arg Pro CCC

FIGURE 8

TTATTGCAACCGTGAGCGCTGGCCACTCACGATCCAGCCCTGCCACTCCGGTCCGGGCAG <u> AATAACGTTGGCACTCGCGACCGGTGAGTGCTAGGTCGGGACGGTGAGGCCAGGCCCGTC</u> 100 90

091. 150 140

240 T C G C A T T C A T C G C C C A A C G G C C A G T G G G C C A A G G T A G G G C T A G G C C T C C T <u> AGCGTAAGTAGCGGGGTTGCCGTCACCCGGTGACCCCCGTTTCATCCCCGATCCGGAGGA</u> S S ے R. G C R H P V 210 200

TAGTGAAGCGTTACGGGTTCTGTTAACGCATGCTGCTTCTCCGGGCAGCGCGGAGCTCG ATCACTICGCAATGGCCAAGACAATTGCGTACGACGAAGAGGCCCGTCGGCCTCGAGG 280 2.70

GGGGCTTGAACGCCCTCGCCGAFGCGGTAAAGGTGACATTGGGCCCCAAGGGCCGCAACG CCCCGAACTTGCGGGAGCGGCTACGCCALTTCCACTGTAACCCGGGGTTCCCGGCG11GC ح 2 G B 340 ALADAVKV

1CG1CC1GGAAAAGAAGIGGGTGCCCCCACGAICACCAACGATGGTG1GTCCA1CGCCA AGCAGGACCTITICTICACCCCACGGGGGTGCTAGTGGTTGCTACCACACAGGTAGCGGT C T D A P · T

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X X I I I V I X I X I X I I I I

(CONT'D)

FIGURE 8

AGGAGATCGAGCTGGAGGATCCGTACGAGAAGATCGGCGCCGAGCTGGTCAAAGAGGTAG TCCTCTAGCTCGACCTCCTAGGCATGCTCTTCTAGCCGCGGGCTCGACCAGTTTCTCCATC I G A 400 ¥ 450 >

CCAAGAAGACCGATGACGTCGCCGGTGACGGCACCACGACGGCCACCGTGCTGGCCAGG GGTICITCIGGCIACIGCAGCGGCCACIGCCGIGGIGCIGCCGGIGGCACGACCGGGICC 630 G 0 J > ۵

000 CGTTGGTTCGCGAGGCCTGCGCAACGTCGCGGCCGGCGCCAACCCGGCTCGGTCTCAAAC GCAACCAAGCGCTCCCGGACGCGTTGCAGCGCCGGCCGCGGTTGGGCGAGCCAGAGTTTG 590 ے z ک ن 580 ⋖ > Z G ш

GCGGCATCGAAAAGGCCGTGGAGAAGGTCACCGAGACCCTGCTCAAGGGCGCCAAGGAGG CGCCGTAGCTTTTCCGGCACCTCTTCCAGTGGCTCTGGGACGAGTTCCCGGGGTTCCTCC 650 840 >

V E T K E Q I A A T A A I S A G D Q S I TCGAGACCAAGGAGCAGATTGCGGCCACCGCAGTTTCGGCGGGTGACCAGTCCATCG **AGCTCTGGTTCCTCGTCTAACGCCGGTGGCGTCGCTAAAGCCGCCCACTGGTCAGGTAGC** 700 000

GTGACCTGATCGCCGAGGCGATGGACAAGGTGGGCAACGAGGGCGTCATCACCGTCGAGG CACTGGACTAGCGGCTCCGCTACCTGTTCCACCCGTTGCTCCCGGCAGTAGTGGCAGCTCC ی ш Z > ≚ ے ع

FIGURE 8 (CONT'D)

040 **TCAGGTTGTGGAAACCCGACGTCGAGCTCGAGTGGCTCCCATACGCCAAGCTGTTCCCGA** <u> AGTCCAACACCTTTGGGCTGCAGCTCGAGCTCACCGAGGGTATGCGGTTCGACAAGGGCT</u> 810 800 G 790

TGTAGAGCCCCATGAAGCACTGGCTGGGCCTCGCAGTCCTCCGCCAGGACCTCCTGGGGA ⋖ C E S ے 000

TGTAGGACGACCAGTCGAGGTTCCACAGGTGACAGTTCCTAGACGACGGCGACGAGCTCT **ACATCCTGCTGGTCAGCTCCAAGGTGTCCACTGTCAAGGATCTGCTGCCGCTGCTGCTCGAGA** 940 S s × ഗ

T @ C A GT A G C C T C G G C C A T T C G G C G A C G A C T A G T A G C A G C T C C T G C A G C T C C G C T C C <u> AGGTCATCGGAGCCGGTAAGCCGCTGCTGATCATCGCCGAGGACGTCGAGGGCGAGGCGCG</u> G 1000 G

1080 TGTCCACCCTGGTCGTCAACAAGATCCGCGGCACCTTCAAGTCGGTGGCGGTCAAGGCTC S 1000 **5** ≃ 1040

GGCCGAAGCCGCTGGCGGCGTTCCGCTACGACGTCCTATACCGGTAAGAGTGGCCACCAG C C G G C T T C G G C G C C G C C A A G G C G A T G C T A C G G C C A T T C T C A C C G G T G G T C Ξ <u>_</u>

T C C A C T A G T C G C C T C C C G G A C T G C G A C C T C T I G C G G C T G G A C A G C G A C C G T I GGAGAAC GCCGACCTGTCGCTGCTAGGCA <u> AGGTGATCAGCGAAGAGGTCGGCCTGACGCT</u> S

(CONT'D

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CGGTGGTGTCGCGGTGA

CCGACGCCATCGCCGGACGAGTGGCCCAGATCCGCCAGGAGATCGAGAACAGCGACTCCG GGCTGCGGTAGCGGCCTGCTCACCGGGTCTAGGCGGTCCTCTAGCTCTTGTCGCTGAGGC **TCCGGGCGTTCCAGCACCAGTGGTTCCTGCTCTGGTGGTAGCAGCTCCCGGCGACTGT** 1300 d J ~ 0

1380 1330

TGATGCTGGCACTCTTCGACGTCCTCGCCGACCGGTTCGACCGGCCACACAGCGCCACT

ACTACGACCGTGAGAAGCTGCAGGAGCGGCTGGCCAAGCTGGC

C

1500 GCCAAGCGTTACBBTTCCBGCGGCAGCTCCTCCCGTAGCAGCGGCCACCCCCACACTGCG 480 ى ш CGGTTCGCAATGCCAAGGCC

1560 | CGAGCTTCCGCTGCTCCGCTGGCCGC CGAAGGCGACGAGGCGA GGTTGTAGCACTTCCACCGCGACCTCCGGGGGGAC **ACAACGTTCGCCGGGGCTGGGACCT** CCAACATCGTGAAGGTGGCGC1

FIGURE 8 (CONT'D)

z	INCOTINGTINGCONANDANG TOCOCANCOTOCONOCINACOTONOCACONOCINACO	CGCACCACCGGCTCTTCCACGCGTTGGACGGCCGACCGGTGCCTGACTTGC	1680
_	CIG	GAC	
င	CGA	CCT	
=	CAC	GTG	1670
G	CGC	500	
P A G II G	CCT	CGA	
٩	ככפ	ესს	1660
_	CTG	GAC	16
Z	AAC	116	
~	CGC	CCG	9
>	GTG	CAC	1850
V V A E K V R N L	AAG	TTC	
Ш	GAG	CTC	
<	CCC	CGG	1640
>	GTG	CAC	_
>	GTG	CAC	
ۍ	CCC	000	30
ط	500	SSS	18
ш	TGGAGCCGGG	ACCTCGGCCC	
_	ŢG	AC	

GAGTCTGGCCACAGATGCTCCTAGACGAGGGACGGCCGCAACGACTGGGCCAGTTCCACT **CTCAGACCGGTGTCTACGAGGATCTGCTCGCTGCCGGCGTTGCTGACCCGGGTCAAGGTGA** 1700 1690

1800 T R S A L Q N A A S I A G L F L T T E A CCCGTTCGGCGCTGCAGAATGCGGCGTCCATCGCGGGGCTGTTCCTGACCACCGAGGCCG 1790 1780 1760

TCGTTGCCGACAAGCGGGAAAAGGAAAGGCTTCCGTTCCCGGTGGCGGCGACATGGGTG 1880 IJ C 1840 > S ⋖ 1030 ш 1820 ш ۵ ۵ 1810

C C C G A G G A G C C A C G A T G C C A T G G C T C T T G T G G T C C G T C C G T T G G A A GGGCTCCTCTGGTTGGGAGCTACGGTACCGAGAACACCCACGCAGTCGTGTAGGCAACCT] 1970 1960 1950 1940 1930

2040 <u> ACCGGCGACACCCGCTCAGCCCCCGGCGCAGAGCCACGTCGTCGCGCGCCTACCCATGCT</u> 2000 8 (CONT'D)

FIGURE

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... 2400

2390

2380

2370

2360

2350

GTTTGAGGTCTACCTTTTTGGCACCCACGGATTCGAGGATAGGCGCCGATGTTTACTC CAAACTCCAGGATTCGAGGATTCGAGGCTACCCCGATGTTTACTC CAAACTCCAGGATGCGTGCCTAAGCTCCTATCCGCGGCTACAATGAG 2410 2410 2410 2410 2410 2410 2410 241
GTTTGAGGTCTACCTTTTTGGCACCCACGGATTCGAGGATAGGCGCCCGATGTGCAAACTCCAGGATTCGAGGATAGGCGCCCGATGTGCAAACTCCAGGATTCGAGGATAGGCGCCCGATGTGTGCTCCTATCCGCGGCTACAGGCTACAGGCTACAGGCGCTACAGGCGCTACAGGCGCTACGGCTAGGCGCTAGGCGCCAGGCGCTGGCTAGGCGCTAAGCGCCAGGCGCTGGCTTGGCTGGC
GTTTGAGGTCTACCTTTTTGGCACCCACGGATTCGAGGATAGGCCAAAACCGTGGGTGCCTAAGCTCCTATCCCTATCCCTATCCCTATCCCTATCCCTATCCCTATCCCTATCCCTATCCCTATCCCCGACCGA
GTTTGAGGTCTACCTTTTTGGCACCCACGGATTCCAAACCGTGGCTGCCTAACCAAACCGTGGGTGCCTAACCGAACCGTGGGTGCCTAACCGAACCGACGGGTGCCTAACCGAACCGGGCTAGGCGAACCGGAAGCGGGATGCCGGAACCGGGCTAGAACCGGAACCGGGATGCCGGAACCGGGATGCCGGAACCGGGATGCCGGAACCGGGATGCCGGAACCGGGATGCCGGAACCGGGATAAAACCGGGAACCCGGGAACCCGGGAACCCGGGAACCCGGGAACCCGGGATAAAAAAAA
GTTTGAGGTCTACCTTTTTGGCA CAAACTCCAGATGGAAAAACCGT 2410 2420 5GAACCGACCGGCTGCCGATCC 3CTTGGCTGCCGACGGGCTAGG S G V P Q G I R 2470 2480
UU ()()

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FIGURE 8 (CONT'D)

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2580	CCGCAAGCCCGAGTCGTTGCTGCCCGAGTTGACGAAGCTCGGGTAGCTGGTGCCAGGGCT	CCGA	R L G S D H S G S H V F S P Y S T G P · S	2640	GGGTTTGCGCCGAGCCAGCCGCGGCACTGCCGCTACCGGGGTTCGGGTT	CCAA	RLGPHAGSGAAAASGSGPNPN	2700	GCCTGAGTEEAGGCCGCCAACAGGAGCACTGGCCGGGGCGGCGACGGGCGTGTTGGTCAG	CGGACTCAGGTCCGGCGGTTGTCCTCGTGACCGGCCCCGCCGCTGCCCGCACAACCAGTC	_
	CAG	GTC	<u>م</u>		100	AGC	<u>a</u>		TGG	ACC	 -
		CG	G		10t	CA	Ξ		LCT	ACA.	エ
2570	[00]	CC!	—	2630	500	CC	ح	2890	.00T	\ <u>\</u> \\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	—
26	100	700	S	26) YC	100	G	26	CCC	000	۵
	GTA	CAT	>-		CCT	CGA	S		GAC	CTG	>
5	SCG	CCC	<u>د</u>	53	CCC	SSS	IJ		CCC	ນນນ	∢
2580	CCT	CGA	S	2820	ACT	TGA	S	2680	CGC	500	∢
	GAA	CTT	u_	_	CCC	500	≺		บบว	CCC	ط
	3AC	CTG	>		CGC	300	∢		300	SCG	∢
2550	111	CAA	=	2610	V GC		⋖	2870	ACT(I GA(S
2	CAC	CTC	S	26	CCC	997	J	26	1001	ַנני.	∢
•		700	J		CAC	CTC	s		755		۵
0	CTC	CAC	S	ව		CCC	J	0	ACA	101	>
2540	116	AAC	Ξ	2800	CC	000	~	2880	CCA	GGT	J
	TCG	AGC	۵		111	AAA	=		000	CCC	ق
	GAG	CTC	S		555	CCC	ے		AGG	TCC	_1
2530)))	3000	(7	2590		UUU	ح	2850	٦٢	466	_
25	AGG	TC	,	28	וטטו	CC	,	26	BAG	TC.	
	CCA	100			TCTAAGGCCC	ATI	- -		CTC	GAC	
	U U	S	~		1	ΑC	=		O U	CG	G

CGAGGGGGGA GGCGAGGATGCCCGAACTCAAAGCCGCCGTGCTCATGCCGCCGGTGGCGTAGCCGGCGGA C C G C T C C T A C G G G C T T G A G T T T C G G C C A C G A G T A C G G C C A C C G C A T C G G C C G C C T GCCCGAGTTGAGGACGTTCGCCAGGCCGTGTTGGAGACCGCCCGTTGATC 2810 J J 2800 2790 ∢ 2780 z S

GCTGACCAAGGCCGCCTCCGAGCCAGCCGCTTCCTAAGGCGGCGTTTTGCATCCCGG **∀** | 2800 2850 2840 2830

2940 GTTCCAGAAGCTGGTGTTGAGGCTGCCTGCCTGCCGAGGCCCGCGTTGATTGTCCCCGA C A A GGT C T T C G A C C A C A C C C G A C G G G C G G G C T C C G G G C G C A C T A A C A G G G G C T 2920 2910 2900

(CONT'D)

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FIGURE

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2980 2970 2980 2990	3000
GGTCCCGATGCCGCTGTTCAGGGAGCCCGAATTCCCGATGCCGATGTTTCCGCTGCCGGA	CTGCCGGA
CCAGGGCTACGGCGACAAGTCCCTCGGGCTTAAGGGCTACGGCTACAAAGGCGCT	CACGGCCT
T G I G S H L S G S H G I G I H G S G S	S G S
3 3020 3030 3040 3050	3060
GTTGAATAAGCCGACGTTGCCGGTGCCCGAGTTCCCGAAGCCGATGTTGCCGCTACCCGA	CTACCCGA
TATT	GATGGGCT
NFLGVNGTGSNGFGINGSGS	S G S
3070 3080 3090 3100	3120
GTTGAAGCCGCCGAAACCCATCTGGTGATCACCGGTGATCCCGGAACCCGATATTCCCGCT	TTCCCGCT
CAACTTCGGCGGTTTGGGTAGACCACTAGTGGCCACTAGGGCTTGGGCTATAAGG	AAGGGCGA
NFGGFGMQHDGT1GFGINGS	S , D 7
3130 3140 3150 3160 3170 3180	3180
ACCGGTGTTGCCGAAGCCGATATTCCCGTCGCCGAGGTTGCCGAGGCCCAGGTTGC	TTGCCGCT

3190 3200 3210 3210 3230 3230 3240 GCCGGTGTTGCCGCTGCCGGTGTTGCCGCTGCCGATGTTGTTGTT CGCCACACGGCGACGCTACAACAACAA G S G I N G T G S G I N H H 3260 3260 3260 3300 GCCGATGTTGTTGTTGTTGTTGTTGTTGCCGCTGCCGCTGCCGGTGTTGCCGAA CGGCTACAACAACAACAACAACAACAACGGCTACAACGGCCACAACGGCTTGCTACAACAACGGCTACAACAACGGCTACAACGGCTACAACGGCTACAACGGCTACAACGGCTACAACGGCTACAACGGCTACAACGGCTACAACGGCTACAACGGCTACAACAACAACAACGGCTACAACAACAACAACAACAACGGCTACAACAACAACAACAACAACAACAACAACAACAACAAC						•			
3196 CGGTGTT GCCACAA(T N 3266 CGATGTT GCTACAA(I N	3240	311611	CAACAA	=======================================	3300	BCCGAA	TLOSS	u U	
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TGGCCACAACGGCTTCGGCTATAAGGGCAGCGGCTCCAACGGCTCCAACGGCGA

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FIGURE

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3370 3380 3390 3400 3400 3420 3420 3470 3410 3420 3420 3420 3420 3410 3420 3420 3420 3410 3420 3420 3420 3420 3420 3420 3420 342	3490 3590 3510 3520 3530 3540 CCCCACATTCGTACCACCACCACCACCACCACCACCACCACCACCACCACC	3580 3580 3670 3590 3590 3690 3600 3600 3600 3600 3600 3600 36	3818 3828 3838 3848 3858 3868 3888 3888 3888 3888 TGCGGCCGCCGCCGCCGCCGCCGCCAGGCTAGGTACTGGGTTGC ACGCCGCCGCCGCCGCCGCCGCCGCCGCCGCCGCGCGCG	3710 3720 3720 3720 3720 3720 3720 3720 CGGCCACTAGTCAGTTCGGATGT CGGCCGCCATCAGTCAGTCAGTCGGATGT CGGCCGGTAGTCAGTCAGTCAGTCGGTCGGTCGGTCGGTC
3370 CCAGGGCGCCA GGTCCCGCGGT W P A L 3430 CACGTCCAATG GTGCAGGTTAC	3490 CTGCCAAAC GACGGGTTTC	36 TGCCGCT ACGGCCGA	TGCGGCCG ACGCCGGC	3878 GACGGCCATC

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FIGURE

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40	TC AG	80 00 00 00 00 00 00 00 00 00 00 00 00 0	0 0 0 0 0 0 0 0	20 00 00 00	4080 GCCC CGGG
38	TYC LAG	39 1AC :TG	9000 0000 0004	40 3CA CGT	40 300 300 300
	TG/	333	GA C	AT TAO	GE
	TAT TAT	CA GT	ITG VAC P	200 200 200 200	2000 1000 1000
30	CGA GCT S	90 TG1 ACA	50 CT1 GA/	110 CCC CCC CCC CCC CCC CCC CCC CCC CCC	4070 CGACG GCTGC R R
38	000 C00 V	38 AC TG	39 1 A T 1 T A	40 66 7 7	48 000 000 000 000 000 000 000 000 000 0
	AGT FCA	TTA AAT	GTC CAC		SAC
	TC/ AG	AC.		CT	AG.
0	VCA FGT	38 CCA 361	40 ITC AAC	88 50 50 70 70	30 A
382	AA/ TTT F	386 AT(TA(39, CT GA, K	4 0 0 0 0 0	4000 1660 ACCG
	3GC CGG	20C	CAT STA	300	4000 407 GTAIGGCAAGCACCGA CATACCGTICGTGGCT Y P L C R
)))))	AG(TC(CT(GA(S	TA ATO	TG(AC(
0	GCT	3860 3870 3880 3890 3 ACGCAAAATGCGGGCTTGTCAGCCGATCCAACTTAACTGTCAGCGA TGCGTTTTACGCCCGAACAGTCGGCTAGGTTGAATTGACAGTCGCT A F II P S T L R D L K V T L S	3Ø CCA GGT	98 369 566	4050 ATGTCGCTG TACAGCGAC
381))))	38 TT(AA(39: TA(3000	4050 GTCG CAGC
	001	300 000 000 8	CAA STT	3TC	IAT ITA
	(00) (00) (00)	200 200 900 900 900	TT(AA(T A (CGTA GCAT Y
30	200	30 ATG TAC	2Ø CAC GTG	30 70 70 70 70 70	200 200 200 200 200 200 200 200 200 200
386	000 CC/	386 AA/ TT	393 666 666 666	398 TT/ AA	4040 TGCCC ACGGG A
	\GC rcG -	SCA	VTC FAG	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	CCT GGA G
	TT/ A.A.	AC(GT.	7	CAC
90	CAA STT	SØ ACC TGG	10 30 10 10 P	78 200 200 A	00 00 00 00 00 00
379	AGC TCC	38.0 CA.	39) TG(AC(397 AA(TT(4030 1010 AGAC
	200 200 8	200 200 200 200	200 200 200	466 700 P	166 100 100
	GGCCGCAGCAATTAGCGGICCCGACCCGGGAACCAACATCAGTGCCGAATTGATCTC CCGGCGTCGTTAATCGCCAGGCTGGGCCCTGGCCGTTTGTAGTCACGGCTTAACTAGAG A A A I L P G S G P G A F M L A S N I E	3850 3860 3870 3880 3990 3900 TGGCGGCAACTTAACTGTCAGCGACGACGACCGACCGACC	3910 3920 3930 3940 3950 3960 TTGCCGTGCGGTATCGGCACTTCAATACCACTCATCTTTGGGGTCATCTTTGGAGCGCCACTCGCGCCCCGGGGGGGG	3970 3980 3990 4000 4000 4010 4020 CCTAGGAACCGCCAGCTTACCTAGTCCCGGGTAGGGCCCGACTGGCGGCCGGGCGGCGGCGGCGGCGGCGGCGGCGGCGGC	4030 4040 4050 4080 4080 4090 4080 A080 A080 A080 A080 A080 A080 A08
	50	⊢ ∢	· - ~	0	⊢ ∢

GCCTCACCGACCAACTCGCCTGCTACCGCCGACCCCCAGCGCCAACAGCATTGCGTGGC CGGAGTGGCTGGTTGAGCGGACGATGGCGGGCTGGGGGTCGCGGGTTGTCGTAACGCACCG AAGAGTIGCICCGCGACGCGTICACCCGGTIGATCGAACATGTCGACGAACTCACCGACG TICICAACGAGGCGCIBCGCAAGIGGGCCAACIAGCIIGIACAGCIGCTIGAGIGGCIGC 4180 4160 4160

4090

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FIGURE 8 (CONT'D)

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